

Judul Artikel: The Role of Nrf2 Transcription Factors in Various Physiological and Pathological States

1. COVER JURNAL



Vol. 6 No. 2 (2023): Acta Biochimica Indonesiana



The articles in this issue are contributed by authors affiliated with 8 institutions from 2 countries, including **Indonesia** (Universitas Indonesia, Universitas Negeri Padang, Universitas Andalas, Universitas Lampung, Universitas Nusa Cendana, and Bandung Institute of Technology) and **India** (Integral University and Prasad Institute of Medical Sciences).

[Cover Download](#)

PUBLISHED: 2023-12-31

[Register](#) [Login](#)

eISSN 2654-3222 / pISSN 2654-6108



ACTA BIOCHIMICA INDONESIANA

JOURNAL OF INDONESIAN SOCIETY FOR BIOCHEMISTRY AND MOLECULAR BIOLOGY

[HOME](#) [ABOUT](#) [ARTICLES](#) [SUBMISSIONS](#) [EDITORIAL TEAM](#) [PUBLICATION ETHICS](#) [CONTACT](#)

Acta Biochimica Indonesia (ActaBiolina) is a peer-reviewed and open-access journal that disseminates original research articles and review articles covering diverse Biochemistry and Molecular Biology topics. The journal is published biannually by the Indonesian Society for Biochemistry and Molecular Biology.

[Find more information about the scope and aim here.](#)



Benefits of publishing with Acta Biochimica Indonesia

Speed of publication

Acta Biochimica Indonesia offers a fast publication schedule whilst maintaining rigorous peer review; all articles must be submitted online, and peer review is managed fully electronically. Articles will be published with

[Submission](#)
[Article Template \(Doc\)](#)
[Author Guidelines](#)
[Editorial Team](#)
[Reviewer](#)

[Register](#) [Login](#)

eISSN 2654-3222 / pISSN 2654-6108



ACTA BIOCHIMICA INDONESIANA

JOURNAL OF INDONESIAN SOCIETY FOR BIOCHEMISTRY AND MOLECULAR BIOLOGY

[HOME](#) [ABOUT](#) [ARTICLES](#) [SUBMISSIONS](#) [EDITORIAL TEAM](#) [PUBLICATION ETHICS](#) [CONTACT](#)

[HOME](#) / [ARCHIVES](#) / VOL. 6 NO. 2 (2023): ACTA BIOCHIMICA INDONESIANA / Reviews

The role of Nrf2 transcription factors in various physiological and pathological states

Deasyka Yastani
 Department of Biochemistry, Faculty of Medicine, Universitas Indonesia,
 Jakarta 10430, Indonesia
<https://orcid.org/0009-0001-0378-5101>

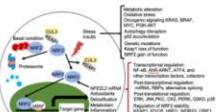


Diagram illustrating the Nrf2 signaling pathway and its role in various physiological and pathological states. The pathway shows Nrf2 being activated by various stimuli (Oxidative stress, DNA damage, Antioxidant response elements, Keap1, etc.) which leads to the recruitment of coactivators (p65, RelB, RelA, p50, etc.) and transcription factors (ATF4, Nrf2, etc.) to the promoter region. This results in the expression of target genes (Phase II enzymes, Antioxidants, etc.) that protect against further damage.

[Submission](#)
[Article Template \(Doc\)](#)
[Author Guidelines](#)
[Editorial Team](#)
[Reviewer](#)

2. DAFTAR ISI DAN ARTIKEL YANG DIAJUKAN UNTUK PENGHARGAAN

Acta Biochimica Indonesiana

<https://doi.org/10.32889/actabioina.103>

REVIEW ARTICLE

Open Access

The role of Nrf2 transcription factors in various physiological and pathological states



Deasyka Yastani¹ , Sri Widia A. Jusman²

¹Department of Biochemistry & Molecular Biology, Faculty of Medicine, Universitas Trisakti, Jakarta, Indonesia

²Department of Biochemistry, Faculty of Medicine, Universitas Indonesia, Jakarta 10430, Indonesia

³Center of Hypoxia and Oxidative Stress Studies, Faculty of Medicine Universitas Indonesia, Jakarta 10430, Indonesia

*Corresponding author: deasyka@trisakti.ac.id

ABSTRACT

Since its first report in 1994, understanding of the nuclear factor erythroid 2-related factor 2 (Nrf2) has continued to grow. Initially recognized for its role in cellular response to oxidative stress, Nrf2 is now known to be involved in a variety of regulatory processes, including metabolic regulation, autophagy, protein homeostasis (proteostasis), and mitochondrial biogenesis. The expression of Nrf2 target genes is highly dependent on stimulus activation and interactions with transcription factors, activators, and repressors. Nrf2 activation serves as a defense mechanism under physiological conditions, but in the context of cancer, it can trigger the development of cancer cells due to its complex roles. Nrf2 is strongly associated with the onset and development of many diseases, including those caused by metabolic disorders and inflammation. Understanding Nrf2's diverse functions offers valuable insights into disease pathogenesis and potential therapeutic approaches. This review explores the pleiotropic role of Nrf2 regulation.

Keywords: Nrf2, oxidative stress, metabolic regulation, proteostasis, transcription factor

Introduction

As a transcription factor, Nrf2 plays a major role in xenobiotic metabolism and the response to reactive oxygen species (ROS) [1,2]. Oxidative stress activates antioxidant genes via the antioxidant response element (ARE) [3,4]. Nrf2 was first reported in 1994 as an NF-E2-like transcription activator binding to the NF-E2/AP1 tandem repeat of the beta-globin locus control region [5,6], but its biological function is not yet clearly known. In 1996, studies in Nrf2 knockout mice revealed no aberrant phenotype, although the mice were more susceptible to stress [7]. In 1997, a study showed that knocking out enzymes such as glutathione S-transferase (GST) and NAD(P)H:quinone oxidoreductase-1 (NQO1), which are regulated by ARE, revealed that Nrf2 controls drug metabolism enzymes in vivo [8]. Recent findings revealed that Nrf2 regulates various processes through ARE and electrophile response elements (EpRE), which play a role in regulating

endogenous responses to various intrinsic and extrinsic stresses [9–15].

Recent studies have further identified new roles of Nrf2 beyond its response to oxidative stress. Nrf2 is involved in regulating inflammation, autophagy, metabolism, protein homeostasis (proteostasis) and unfolded protein response (UPR), and mitochondrial biogenesis, as well as being associated with carcinogenesis [1,2,15,16]. These findings suggest that Nrf2 is a pleiotropic transcription factor. This review presents the role of Nrf2 in various processes inside the cell.

Structure of Nrf2

As a part of family of Cap'n'collar (CNC) proteins, Nrf2 has a unique CNC domain that binds to the basic leucine zipper (CNC-bZIP) motif [2,3]. The Nrf2 protein comprises seven functional domains, known as Nrf2-ECH homology domains (Neh1–

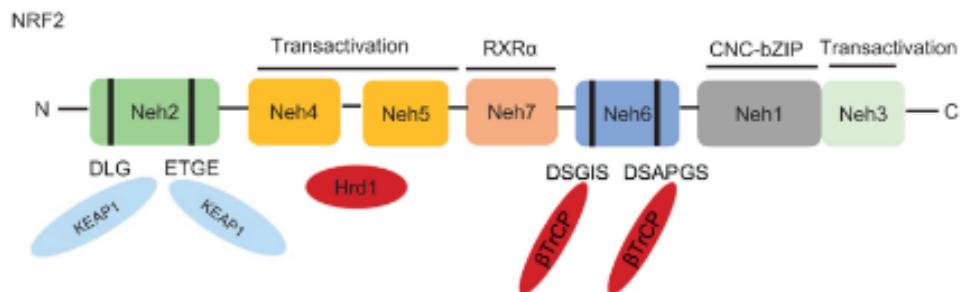


Figure 1. The structure of Nrf2. Nrf2 comprises seven domains, known as Nrf2-ECH homology domains (Neh1- Neh7)

Neh7), each with different functions in regulating Nrf2 transcription activity.

- Neh1 domain: comprises the CNC-bZIP region, enabling heterodimerization with the small musculoaponeurotic fibrosarcoma (sMAF) oncogenes. This interaction is crucial for recognizing ARE and/or EpRE. The CNC-bZIP region is conserved across various species, highlighting its importance for Nrf2 function [15].
- Neh2 domain: Features an N-terminal domain that binds to the negative regulatory domain of Kelch-like ECH-associated protein 1 (KEAP1). This domain contains ETGE and DLG motif, mainly interacting with the domain (KEAP1) for ubiquitination and subsequent Nrf2 degradation [1,2].
- Neh3-5 domains: Serves as domains for transcriptional activation by interacting with transcriptional components [1,2].
- Neh6 domain: Involved in the degradation of Nrf2 independent of Keap1. It contains two redox-independent degrons, DSGIS and DSAPGS, which link to E3 ubiquitin ligase β-transducin repeat-containing protein (βTrCP) [1,2].
- Neh7 domain: Functions to suppress Nrf2 activity by interacting with retinoic X receptor alpha (RXRa).

All of these domains maintain the stability of Nrf2 and facilitate transcriptional activation of the target genes at a stratified level, encompassing transcriptional, post-transcriptional, and post-translational regulation in response to various

stresses [1]. The structure of Nrf2 is illustrated in Figure 1.

Regulation of Nrf2

The regulation of Nrf2 generally occurs through the various stages shown in Figure 2 below.

Regulation of Nrf2 stability

Under basal conditions, Nrf2 is maintained at low levels within cells due to proteasomal degradation mediated by KEAP1 [1,17]. Nrf2 stability is strictly regulated through ubiquitination mediated by E3 ubiquitin ligase complexes and subsequent proteasomal degradation. The ubiquitin ligase complexes involved in this regulation include HRD1/Synoviolin, WD Repeat protein (WDR), βTrCP-S-phase kinase-associated protein-1 (SKP1)-CUL1-RBX1, KEAP1-Cullin (CUL) 3-RING-box protein (RBX)1, and 3-CUL4-damaged DNA binding protein (DDB)1. Each of these complexes plays a role under different cellular conditions [1].

Regulation of Nrf2 transcription

Nrf2 transcription is regulated through various mechanisms involving NF-κB binding sites, the aryl hydrocarbon receptor (AHR)-AHR nuclear translocator (ARNT) complex, other transcription factors, and cofactors [1]. The Nrf2 transcription factor binds to the ARE promoter region to activate gene transcription. The NFE2L2 gene contains the ARE promoter region, a xenobiotic responsive element (XRE), two additional promoters with XRE-like elements, and

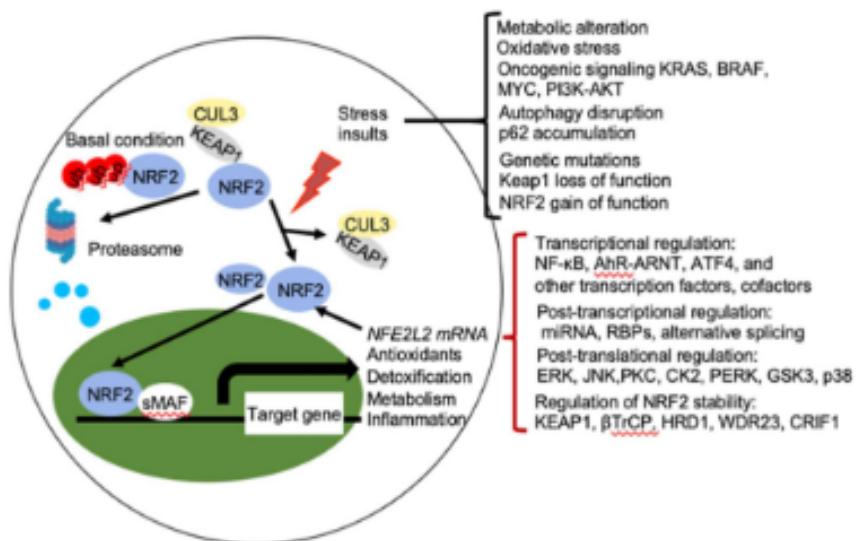


Figure 2. Stages of Nrf2 regulation. Nrf2 regulation involves multiple stages: stability, transcription, post-transcription, post-translation, metabolic changes, response to oxidative stress, oncogene signaling, autophagy regulation, and genetic mutations [1]

NF-κB binding site. The ARE region allows Nrf2 to directly activate the transcription process through a positive feedback mechanism that amplifies Nrf2's effect [1]. Additionally, transcription of NFE2L2 is regulated by various transcription factors, such as NF-κB and AHR. The AHR, in particular, activates the XRE [1,10].

Regulation of Nrf2 post-transcription

Post-transcription regulation of Nrf2 involves microRNAs (miRNAs), RNA-binding proteins (RBPs), and alternative splicing. MicroRNAs are non-coding RNAs, typically 22 nucleotides long, that can suppress gene expression by binding to mRNA molecules in a sequence-specific manner, thereby inhibiting protein translation and destabilizing mRNA. A decrease in Nrf2 transcription activity has been associated with an increase in miR-93, miR-144, miR-153, miR-28, and miR-34. However, the role of miRNAs regulation requires further study and validation [1,18].

Regulation of target gene activation

Nrf2 activation is strictly regulated, as only specific genes can interact with Nrf2 and its binding

component (transcription factors, mediators, and cofactors) to achieve activation. Key transcription factors involved include sMAF transcription factors, BTB and CNC homology transcription factors 1 (BACH1), activating transcription factor 4 (ATF4), activator protein 1 (AP-1) subunit c-Jun, CREB-binding protein (CBP), p300 cofactors, activating transcription factor 3 (ATF3), and other co-activators [1,19].

Regulation of Nrf2 In oxidative stress response and drug detoxification

Nrf2 regulation oxidative stress response and drug detoxification involves the expression of cytoprotective genes that facilitates the synthesis of enzymes involved in drug detoxification and maintenance of cellular hemostasis [1-3].

- Phase I enzymes:** These enzymes participate in reduction, oxidation, and hydrolytic reactions of xenobiotics. Examples include aldehyde dehydrogenase 1 (ALDH1), NQO1, aldo-keto reductases (AKRs), carbonyl reductases (CBRs), and some cytochrome P450 oxidoreductases (CYPs) [3].
- Phase II enzymes:** These enzymes are involved

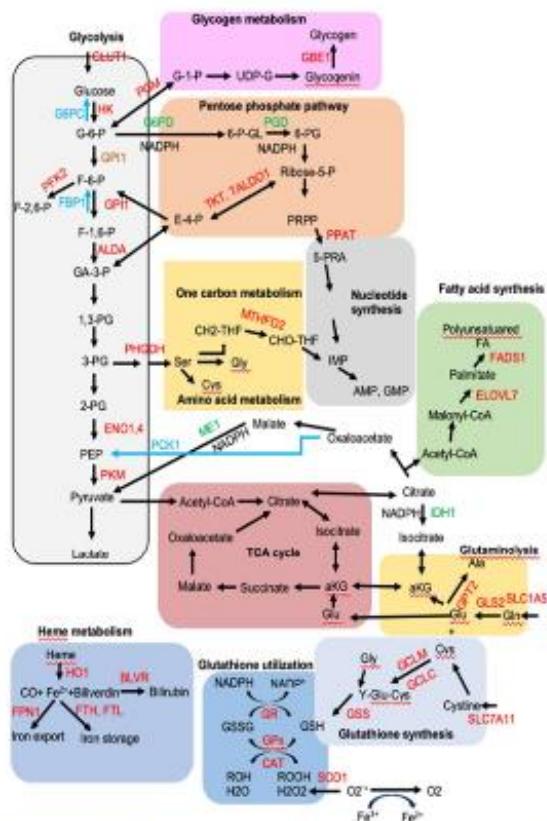


Figure 3. Regulation of Nrf2 on metabolism. Nrf2 enhances the expression of genes involved in glycolysis, glutaminolysis, glycogen metabolism, pentose phosphate pathway, amino acid metabolism, fatty acid synthesis, heme metabolism, glutathione synthesis, and nucleotide biosynthesis (red). Nrf2 decreases the expression of genes associated with gluconeogenesis (blue). Enzymes involved in NADPH production are highlighted in green [1].

in conjugation reactions, such as UDP-glucuronosyltransferase (UGT), glutathione S-transferase (GST), heme oxygenase-1 (HO-1), and UDP-glucuronate synthase [3,20].

c. Phase II enzymes: These drug transporters transport conjugated metabolites after phase II. Example include breast cancer resistant protein (BCRP), ATP-binding cassette G5 (ABCG5), multidrug resistance-associated proteins (MDR), and ATP-binding cassette G8 (ABCG8) [3].

Nrf2-targeted enzymes for ROS elimination include thioredoxin reductase, thioredoxin, catalase (CAT), superoxide dismutase 1 (SOD1), peroxiredoxin, glutathione peroxidase, and sulfiredoxine, as well

as some glutathione S-transferases [2,3,20]. The cytoprotective proteins encoded by Nrf2 are essential for protective mechanisms against various toxins and oxidants, particularly in diseases based on oxidative stress, such as cancer, coronary disease, metabolic syndrome, neurodegenerative diseases, and autoimmune diseases [2].

Regulation of Nrf2 on metabolism

Nrf2 improves the gene expression involved in glycolysis, the pentose phosphate pathways, glycogen metabolism, amino acid metabolism, glutaminolysis, fatty acid synthesis, heme metabolism, nucleotide biosynthesis, and glutathione synthesis. Conversely,

Nrf2 decreases the gene expression correlated with gluconeogenesis. Many studies support that Nrf2 is a major key in the reprogramming of cancer metabolism [2]. The regulatory mechanisms of Nrf2 are illustrated in Figure 3.

Nrf2 regulation on unfolded protein response (UPR) and proteostasis

Misfolded protein accumulation triggers the production of ROS from the mitochondria and endoplasmic reticulum (ER), leading to an increase in activation of Nrf2 [17]. Nrf2 activates transcription of ATF4, which is related to amino acid metabolism. In *C. elegans*, the homologous Nrf2 protein SKN-1 activates various UPR target genes, including ATF6 and XBP1, thereby enhancing the UPR program by maintaining ER integrity and protein homeostasis (proteostasis) [21]. Additionally, Nrf2 promotes the expression of proteasome maturation proteins (POMP), which is crucial for proteostasis [22]. In summary, Nrf2 increases proteasome activity and antioxidants expression, aiding in stress adaptation.

Nrf2 regulation on autophagy

Nrf2 plays a significant role in autophagy by upregulating the expression of autophagy-related genes, including SQSTM1/P62, autophagy protein 5 (ATG5), calcium-binding and coiled-coil domain-containing protein 2 (CALCOCO2/NDP52), gamma-aminobutyric acid receptor-associated protein-like 1 (GABARAPL1), and unc-51-like kinase 1 (ULK1) [17]. Interestingly, the activation of Nrf2 can be stimulated by the accumulation of oxidized proteins or organelles due to impaired autophagy. This increase in Nrf2 is mediated by the accumulation of p62, a multifunctional receptor that stabilizes KEAP 1 and Nrf2, under autophagy-deficient conditions. In conclusion, a positive feedback loop by p62 and Nrf2 can regulate numerous cellular functions [23].

NRF2 regulation on mitochondrial biogenesis

Nrf2 is involved in enhancing the production and growth of new mitochondria within cells. This

process is known as mitochondrial biogenesis. Nrf2 stimulates mitochondrial biogenesis by activating nuclear respiratory factor-1 (NRF-1), is a key regulator of mitochondrial transcription factor A (TFAM) and mitochondrial transcription factor B2 (TFBM2) [24]. TFAM is involved in the transcription and maintenance of mitochondrial DNA, while TFBM2 works alongside TFAM to regulate mitochondrial gene expression. Nrf2 also directly regulates the expression of peroxisome proliferator-activated receptor gamma co-activator 1 alpha (PGC-1 α), a principal regulator of mitochondrial function and biogenesis. The cooperation between Nrf2 and PGC-1 α regulates various aspects of mitochondrial function [24–26].

Conclusion

Nrf2 is a widely expressed transcription factor in mammalian cells, belonging to the Cap'n'collar (CNC) protein family. It features a unique CNC domain that binds to the basic leucine zipper (CNC-bZIP) motif. The Nrf2 protein contains seven functional domains, known as Nrf2-ECH homology domains (Neh1-Neh7). Nrf2 regulation occurs at multiple stages, including stability, transcription, post-transcription, post-translation, and target gene activation. This regulation is highly selective, with only specific genes capable of interacting with Nrf2 and its binding partners to achieve activation. Recent studies have highlighted the pleiotropic nature of Nrf2, demonstrating its roles not only in oxidative stress response and drug detoxification but also in metabolism, heme synthesis, proteostasis, unfolded protein response (UPR), autophagy, and mitochondrial biogenesis.

Acknowledgment

None.

Funding

This review received no external funding.

Declaration of Interest

The authors declare no conflict of interest.

Author contributions

DY: Conceptualization, Writing – original draft; SWAJ: Supervision, Writing – review and editing.

Received: August 30, 2022

Revised: April 20, 2024

Accepted: June 30, 2024

Published online: July 1, 2024

References

- He F, Ru X, Wen T. NRF2, a transcription factor for stress response and beyond. *Int J Mol Sci.* 2020;21(13):1-23. <https://doi.org/10.3390/ijms21134777>
- He F, Antonucci L, Karin M. NRF2 as a regulator of cell metabolism and inflammation in cancer. *Carcinogenesis.* 2020;41(4):405-16. <https://doi.org/10.1093/carcin/bgaa039>
- Hayes JD, Dinkova-Kostova AT, Raghunath A, Sundarraj K, Nagarajan R, Arfuso F, et al. NRF2 regulatory network provides an interface between redox and intermediary metabolism. *Trends Biochem Sci.* 2014;39(4):199-218. <https://doi.org/10.1016/j.tibs.2014.02.002>
- Rushmore TH, Morton MR, Pickett CB. The antioxidant responsive element: Activation by oxidative stress and identification of the DNA consensus sequence required for functional activity. *J Biol Chem.* 1991;266(18):11632-9. [https://doi.org/10.1016/S0021-9258\(18\)99004-6](https://doi.org/10.1016/S0021-9258(18)99004-6)
- Moi P, Chan K, Asunis I, Cao A, Kan YW. Isolation of NF-E2-related factor 2 (Nrf2), a NF-E2-like basic leucine zipper transcriptional activator that binds to the tandem NF-E2/AP1 repeat of the β -globin locus control region. *Proc Natl Acad Sci U S A.* 1994;91(21):9926-30. <https://doi.org/10.1073/pnas.91.21.9926>
- Raghunath A, Sundarraj K, Nagarajan R, Arfuso F, Bian J, Kumar AP, et al. Antioxidant response elements: Discovery, classes, regulation and potential applications. *Redox Biol.* 2018; 17(May): 297-314. <https://doi.org/10.1016/j.redox.2018.05.002>
- Chan K, Lu R, Chang JC, Kan YW. NRF2, a member of the NFE2 family of transcription factors, is not essential for murine erythropoiesis, growth, and development. *Proc Natl Acad Sci U S A.* 1996;93(24):13943-8. <https://doi.org/10.1073/pnas.93.24.13943>
- Itoh K, Chiba T, Takahashi S, Ishii T, Igarashi K, Katoh Y, et al. An Nrf2/small Maf heterodimer mediates the induction of phase II detoxifying enzyme genes through antioxidant response elements. *Biochem Biophys Res Commun.* 1997;236(2):313-22. <https://doi.org/10.1006/bbrc.1997.6943>
- Ma Q. Role of Nrf2 in oxidative stress and toxicity. *Annu Rev Pharmacol Toxicol.* 2013;53(1):401-26. <https://doi.org/10.1146/annurev-pharmtox-011112-140320>
- Suzuki T, Shibata T, Takaya K, Shiraishi K, Kohno T, Kunitoh H, et al. Regulatory Nexus of Synthesis and Degradation Deciphers Cellular Nrf2 Expression Levels. *Mol Cell Biol.* 2013;33(12):2402-12. <https://doi.org/10.1128/MCB.00065-13>
- Tonelli C, Chio IIC, Tuveson DA. Transcriptional Regulation by Nrf2. *Antioxidants Redox Signal.* 2018;29(17):1727-45. <https://doi.org/10.1089/ars.2017.7342>
- Taguchi K, Fujikawa N, Komatsu M, Ishii T, Unno M, Akaike T, et al. Keap1 degradation by autophagy for the maintenance of redox homeostasis. *Proc Natl Acad Sci U S A.* 2012;109(34):13561-6. <https://doi.org/10.1073/pnas.1121572109>
- Baird L, Yamamoto M. The molecular mechanisms regulating the KEAP1- NRF2 pathway. *Mol Cell Biol.* 2020; 40(13):1-23. <https://doi.org/10.1128/MCB.00099-20>
- Ikehata H, Yamamoto M. Roles of the KEAP1-NRF2 system in mammalian skin exposed to UV radiation. *Toxicol Appl Pharmacol.* 2018;360(July):69-77. <https://doi.org/10.1016/j.taap.2018.09.038>
- He F, Antonucci L, Yamachika S, Zhang Z, Taniguchi K, Umemura A, et al. NRF2 activates growth factor genes and downstream AKT signaling to induce mouse and human hepatomegaly. *J Hepatol.* 2020;72:1182-1195. <https://doi.org/10.1016/j.jhep.2020.01.023>
- Mitsuishi Y, Taguchi K, Kawatani Y, Shibata T, Nukiwa T, Aburatani H, et al. Nrf2 Redirects Glucose and Glutamine into Anabolic Pathways in Metabolic Reprogramming. *Cancer Cell.* 2012;22(1):66-79. <https://doi.org/10.1016/j.ccr.2012.05.016>
- Canning P, Sorrell FJ, Bullock AN. Structural basis of Keap1 interactions with Nrf2. *Free Radic Biol Med.* 2015;88(Part B):101-7. <https://doi.org/10.1016/j.freeradbiomed.2015.05.034>
- Hentze MW, Castello A, Schwarzl T, Preiss T. A brave new world of RNA-binding proteins. *Nat Rev Mol Cell Biol.* 2018;19(5):327-41. <https://doi.org/10.1038/nrm.2017.130>
- Wu KC, Cui JY, Klaassen CD. Effect of graded nrf2 activation on phase-i and -ii drug metabolizing enzymes and transporters in mouse liver. *PLoS One.* 2012;7(7). <https://doi.org/10.1371/journal.pone.0039006>
- Fu J, Xiong Z, Huang C, Li J, Yang W, Han Y, et al. Hyperactivity of the transcription factor Nrf2 causes metabolic reprogramming in mouse esophagus. *J Biol Chem.* 2019;294(1):327-40. <https://doi.org/10.1074/jbc.RA118.005963>
- Jang J, Wang Y, Kim HS, Lalli MA, Kosik KS. Nrf2, a regulator of the proteasome, controls self-renewal and pluripotency in human embryonic stem cells. *Stem Cells.* 2014;32(10):2616-25. <https://doi.org/10.1002/stem.1764>
- Pajares M, Cuadrado A, Rojo AI. Modulation of proteostasis by transcription factor NRF2 and impact in neurodegenerative diseases. *Redox Biol.* 2017;11(January):543-53. <https://doi.org/10.1016/j.redox.2017.01.006>

23. Urano A, Furusawa Y, Yagishita Y, Fukutomi T, Muramatsu H, Negishi T, et al. The Keap1-Nrf2 System Prevents Onset of Diabetes Mellitus. *Mol Cell Biol*. 2013;33(15):2996-3010. <https://doi.org/10.1128/MCB.00225-13>
24. Gureev AP, Shaforostova EA, Popov VN. Regulation of mitochondrial biogenesis as a way for active longevity: Interaction between the Nrf2 and PGC-1 α signaling pathways. *Front Genet*. 2019; 10(May): 1-12. <https://doi.org/10.3389/fgene.2019.00435>
25. Piantadosi CA, Carraway MS, Babiker A, Suliman HB. Heme oxygenase-1 regulates cardiac mitochondrial biogenesis via nrf2-mediated transcriptional control of nuclear respiratory factor-1. *Circ Res*. 2008;103(11):1232-40. <https://doi.org/10.1161/01.RES.0000338597.71702.ad>
26. Hu Q, Ren J, Li G, Wu J, Wu X, Wang G, et al. The mitochondrially targeted antioxidant MitoQ protects the intestinal barrier by ameliorating mitochondrial DNA damage via the Nrf2/ARE signaling pathway. *Cell Death Dis*. 2018;9(3). <https://doi.org/10.1038/s41419-018-0436-x>

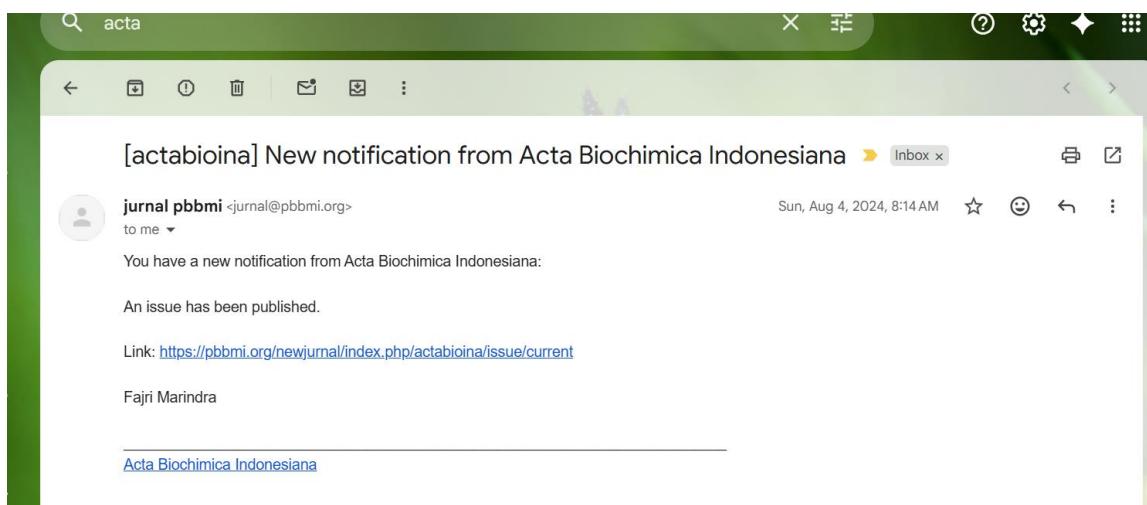
5. BUKTI INDEKSASI DAN KATEGORI JURNAL



The screenshot shows the homepage of the Acta Biochimica Indonesiana journal. At the top right, there are 'Register' and 'Login' buttons. Below them, the journal's eISSN (2654-3222) and pISSN (2654-6108) are displayed. The journal's logo, 'ACTA BIOCHIMICA INDONESIANA', and its full name, 'JOURNAL OF INDONESIAN SOCIETY FOR BIOCHEMISTRY AND MOLECULAR BIOLOGY', are prominently featured. A navigation bar at the top includes links for 'HOME', 'ABOUT', 'ARTICLES', 'SUBMISSIONS', 'EDITORIAL TEAM', 'PUBLICATION ETHICS', and 'CONTACT'. On the left, a section titled 'Indexing & Abstracting' lists various indexing services: DOAJ, SINTA, Garuda, Google Scholar, Scilit, Dimensions, Crossref, and JournalStories. On the right, a sidebar provides links for 'Submission', 'Article Template (Doc)', 'Author Guidelines', 'Editorial Team', and 'Reviewer'.

6. INFORMASI TAMBAHAN - BUKTI JURNAL SUDAH PUBLISHED

PEMBERITAHUAN VIA EMAIL OLEH JURNAL PBBMI



The screenshot shows an email notification from 'jurnal pbbmi <jurnal@pbbmi.org> to me'. The subject is '[actabioina] New notification from Acta Biochimica Indonesiana'. The email body informs the recipient that a new issue has been published and provides a link: <https://pbbmi.org/newjurnal/index.php/actabioina/issue/current>. The email is timestamped 'Sun, Aug 4, 2024, 8:14 AM'.

SINTA Executive

PUBLICATION

Google Scholars

Search...

Sort By: Year

Page 1 of 1 | Total Records : 2

The role of Nrf2 transcription factors in various physiological and pathological states
Author : D Yastani, SWA Jusman
Acta Biochimica Indonesiana 6 (2), 103-103, 2023

The Role of Cytoglobin in Cancer
Author : D Yastani, SWA Jusman
Jurnal Biomedika dan Kesehatan 6 (2), 250-260, 2023

Page 1 of 1 | Total Record 2

Reset Document Req. Synchronization

DEASYKA YASTANI ✓
Sinta ID : 6939388

Date: July, 03rd, 2024

INVOICE

Invoice number: ABI/032/07/2024

Acta Biochimica Indonesiana

E-ISSN 26543222

Indonesian Society for Biochemistry and Molecular Biology

Jl. Salemba Raya No. 06 Jakarta Pusat 10430



Bill To

Deasyka Yastani

Product/ Services	Quantity	Rate/Unit	Total
Article Processing Charge	1	Rp 1.000.000,-	Rp 1.000.000,-
Total Payment			Rp 1.000.000,-

Payment Terms: payment within 7 days

Payment Details:

Money transfer to account below:

Paypal : <https://www.paypal.me/elvirayunita46>

Bank : BNI

Account Number : 0389343728 (Elvira Yunita)

SWIFT Code : BNINIDJAXXX

Payment Confirmation : email to elvirayunita46@gmail.com or 081585183064
(WhatsApp - Elvira Yunita)

Thank you for choosing us as your publication partner

Date: July, 03rd, 2024

INVOICE

Invoice number: ABI/032/07/2024

Acta Biochimica Indonesiana

E-ISSN 26543222

Indonesian Society for Biochemistry and Molecular Biology

Jl. Salemba Raya No. 06 Jakarta Pusat 10430



Bill To

Deasyka Yastani

Product/ Services	Quantity	Rate/Unit	Total
Article Processing Charge	1	Rp 1.000.000,-	Rp 1.000.000,-
		Total Payment	Rp 1.000.000,-

Payment Terms: payment within 7 days

Payment Details:

Money transfer to account below:

Paypal : <https://www.paypal.me/elvirayunita46>
Bank : BNI
Account Number : 0389343728 (Elvira Yunita)
SWIFT Code : BNINIDJAXXX
Payment Confirmation : email to elvirayunita46@gmail.com or 081585183064
(Whatsapp - Elvira Yunita)

Thank you for choosing us as your publication partner

The role of Nrf2 transcription factors in various physiological and pathological states

by dr. Deasyka Yastani M. Biomed PEKERTI2025_02

Submission date: 03-Feb-2026 09:27AM (UTC+0700)

Submission ID: 2869859353

File name: 103-Article_Text-1126-2-10-20240706.pdf (572.77K)

Word count: 3148

Character count: 18401

The role of Nrf2 transcription factors in various physiological and pathological states

Deasyka Yastani¹ , Sri Widia A. Jusman² ¹Department of Biochemistry Molecular Biology, Faculty of Medicine, Universitas Trisakti, Jakarta, Indonesia²Department of Biochemistry, Faculty of Medicine, Universitas Indonesia, Jakarta 10430, Indonesia³Center of Hypoxia and Oxidative Stress Studies, Faculty of Medicine Universitas Indonesia, Jakarta 10430, IndonesiaCorresponding author: deasyka@trisakti.ac.id

ABSTRACT

25

Since its first report in 1994, understanding of the nuclear factor erythroid 2-related factor 2 (Nrf2) has continued to grow. Initially recognized for its role in cellular response to oxidative stress, Nrf2 is now known to be involved in a variety of regulatory processes, including metabolic regulation, autophagy, protein homeostasis (proteostasis), and mitochondrial biogenesis. The expression of Nrf2 target genes is highly dependent on stimulus activation and interactions with transcription factors, activators, and repressors. Nrf2 activation serves as a defense mechanism under physiological conditions, but in the context of cancer, it can trigger the development of cancer cells due to its complex roles. Nrf2 is strongly associated with the onset and development of many diseases, including those caused by metabolic disorders and inflammation. Understanding Nrf2's diverse functions offers valuable insights into disease pathogenesis and potential therapeutic approaches. This review explores the pleiotropic role of Nrf2 regulation.

Keywords: Nrf2, oxidative stress, metabolic regulation, proteostasis, transcription factor

Introduction

As a transcription factor, Nrf2 plays a major role in xenobiotic metabolism and the response to reactive oxygen species (ROS) [1,2]. Oxidative stress activates antioxidant genes via the antioxidant response element (ARE) [3,4]. Nrf2 was first reported in 1994 as an NF-E2-like transcription factor binding to the NF-E2/AP1 tandem repeat of the beta-globin locus control region [5,6], but its biological function is not yet clearly known. In 1996, studies in Nrf2 knockout mice revealed no aberrant phenotype, although the mice were more susceptible to stress [13]. In 1997, a study showed that knocking out enzymes such as glutathione S-transferase (GST) and NAD(P)H:quinone oxidoreductase-1 (NQO1), which are regulated by ARE, revealed that Nrf2 controls drug metabolism enzymes *in vivo* [8]. Recent findings revealed that Nrf2 regulates various processes through ARE and electrophile response elements (EpRE), which play a role in regulating

endogenous responses to various intrinsic and extrinsic stresses [9–15].

Recent studies have further identified new roles of Nrf2 beyond its response to oxidative stress. Nrf2 is involved in regulating inflammation, autophagy, metabolism, protein homeostasis (proteostasis) and unfolded protein response (UPR), and mitochondrial biogenesis, as well as being associated with carcinogenesis [1,2,15,16]. These findings suggest that Nrf2 is a pleiotropic transcription factor. This review presents the role of Nrf2 in various processes inside the cell.

Structure of Nrf2

As a part of family of Cap'n'collar (CNC) proteins, Nrf2 has a unique CNC domain that binds to the basic leucine zipper (CNC-bZIP) motif [2,3]. The Nrf2 protein comprises seven functional domains, known as Nrf2-ECH homology domains (Neh1–

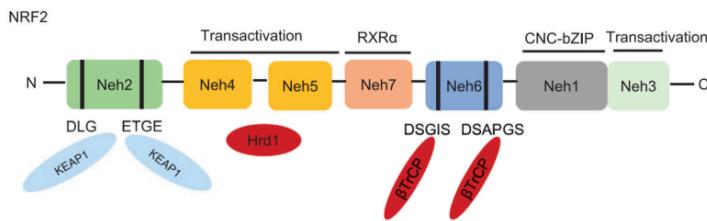


Figure 1. The structure of Nrf2. Nrf2 comprises seven domains, known as Nrf2-ECH homology domains (Neh1- Neh7)

Neh7), each with different functions in regulating Nrf2 transcription activity.

- (a) Neh1 domain: comprises the CNC-bZIP region, enabling heterodimerization with the small musculoaponeurotic fibrosarcoma (sMAF) oncogenes. This interaction is crucial for recognizing ARE and/or EPRE. The CNC-bZIP region is conserved across various species, highlighting its importance for Nrf2 function [15].
- (b) Neh2 domain: Features an N-terminal domain that binds to the negative regulatory domain of Kelch-like ECH-associated protein 1 (KEAP1). This domain contains ETGE and DLG motif, mainly interacting with the domain (KEAP1) for ubiquitination and subsequent Nrf2 degradation [1,2].
- (c) Neh3-5 domains: Serves as domains for transcriptional activation by interacting with transcriptional components [1,2].
- (d) Neh6 domain: Involved in the degradation of Nrf2 independent of Keap1. It contains two redox-independent degrons, DSGIS and DSAPGS, which link to E3 ubiquitin ligase β -transducin repeat-containing protein (β TrCP) [1,2].
- (e) Neh7 domain: Functions to suppress Nrf2 activity by interacting with retinoic X receptor alpha (RXRa).

All of these domains maintain the stability of Nrf2 and facilitate transcriptional activation of the target genes at a stratified level, encompassing transcriptional, post-transcriptional, and post-translational regulation in response to various

stresses [1]. The structure of Nrf2 is illustrated in Figure 1.

Regulation of Nrf2

The regulation of Nrf2 generally occurs through the various stages shown in Figure 2 below.

Regulation of Nrf2 stability

Under basal conditions, Nrf2 is maintained at low levels within cells due to proteasomal degradation mediated by KEAP1 [1,17]. Nrf2 stability is strictly regulated through ubiquitination mediated by E3 ubiquitin ligase complexes and subsequent proteasomal degradation. The ubiquitin ligase complexes involved in this regulation include HRD1/Synoviolin, WD Repeat protein (WDR), β TrCP-S-phase kinase-associated protein-1 (SKP1)-CUL1-RBX1, KEAP1-Cullin (CUL) 3-RING-box protein (RBX)1, and 3-CUL4-damaged DNA binding protein (DDB)1. Each of these complexes plays a role under different cellular conditions [1].

Regulation of Nrf2 transcription

Nrf2 transcription is regulated through various mechanisms involving NF- κ B binding sites, the aryl hydrocarbon receptor (AHR)-AHR nuclear translocator (ARNT) complex, other transcription factors, and cofactors [1]. The Nrf2 transcription factor binds to the ARE promoter region to activate gene transcription. The NFE2L2 gene contains the ARE promoter region, a xenobiotic responsive element (XRE), two additional promoters with XRE-like elements, and

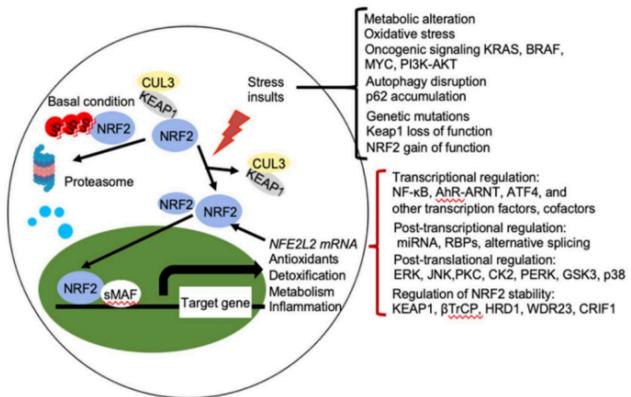


Figure 2. Stages of Nrf2 regulation. Nrf2 regulation involves multiple stages: stability, transcription, post-transcription, post-translation, metabolic changes, response to oxidative stress, oncogene signaling, autophagy regulation, and genetic mutations [1]

NF-κB binding site. The ARE region allows Nrf2 to directly activate the transcription process through a positive feedback mechanism that amplifies Nrf2's effect [1]. Additionally, transcription of NFE2L2 is regulated by various transcription factors, such as NF-κB and AHR. The AHR, in particular, activates the XRE [1,10].

Regulation of Nrf2 post-transcription

Post-transcription regulation of Nrf2 involves microRNAs (miRNAs), RNA-binding proteins (RBPs), and alternative splicing. MicroRNAs are non-coding RNAs, typically 22 nucleotides long that can suppress gene expression by binding to mRNA molecules in a sequence-specific manner, thereby inhibiting protein translation and destabilizing mRNA. A decrease in Nrf2 transcription activity has been associated with an increase in miR-93, miR-144, miR-153, miR-28, and miR-34. However, the role of miRNAs regulation requires further study and validation [1,18].

Regulation of target gene activation

Nrf2 activation is strictly regulated, as only specific genes can interact with Nrf2 and its binding

component (transcription factors, mediators, and cofactors) to achieve activation. Key transcription factors involved include sMAF transcription factors, BTB and ²⁶C homology transcription factors 1 (BACH1), activating transcription factor 4 (ATF4), activator protein 1 (AP-1) subunit c-Jun, CREB-binding protein (CBP), p300 cofactors, activating transcription factor 3 (ATF3), and other co-activators [1,19].

Regulation of Nrf2 in oxidative stress response and drug detoxification

Nrf2 regulation oxidative stress response and drug detoxification involves the expression of cytoprotective genes that facilitates the synthesis of enzymes involved in drug detoxification and maintenance of cellular hemostasis [1-3].

- Phase I enzymes: These enzymes participate in reduction, oxidation, and hydrolytic reactions of xenobiotics. Examples include aldehyde dehydrogenase 1 (ALDH1), NQO1, aldo-keto reductases (AKRs), carbonyl reductases (CBRs), and some cytochrome P450 oxidoreductases (CYPs) [3].
- Phase II enzymes: These enzymes are involved

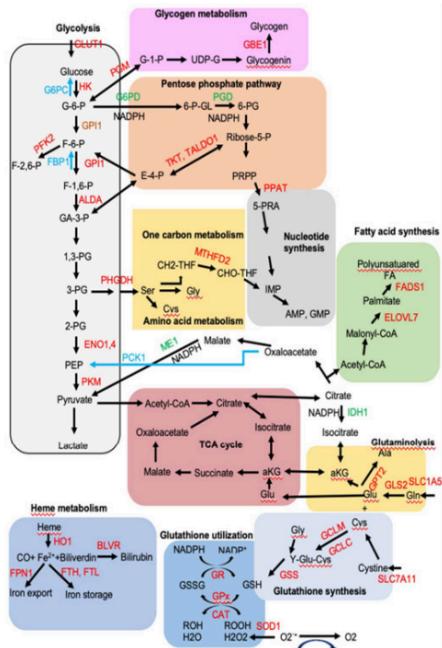


Figure 3. Regulation of Nrf2 on metabolism. Nrf2 enhances the expression of genes involved in glycolysis, glutaminolysis, glycogen metabolism, pentose phosphate pathway, amino acid metabolism, fatty acid synthesis, heme metabolism, glutathione synthesis, and nucleotide biosynthesis (red). Nrf2 decreases the expression of genes associated with gluconeogenesis (blue). Enzymes involved in NADPH production are highlighted in green [1].

in conjugation reactions, such as UDP-glucuronosyltransferase (UGT), glutathione S-transferase (GST), heme oxygenase-1 (HO-1), and UDP-glucurone synthase [3,20].

c. Phase II enzymes: These drug transporters transport conjugated metabolites after phase II. Example include breast cancer resistant protein (BCRP), ATP-binding cassette G5 (ABCG5), multidrug resistance-associated proteins (MDR), and ATP-binding cassette G8 (ABCG8) [3].

Nrf2-targeted enzymes for ROS elimination include thioredoxin reductase, thioredoxin, catalase (CAT), superoxide dismutase 1 (SOD1), peroxiredoxin, glutathione peroxidase, and sulfiredoxine, as well

as some glutathione S-transferases [2,3,20]. The cytoprotective proteins encoded by Nrf2 are essential for protective mechanisms against various toxins and oxidants, particularly in diseases based on oxidative stress, such as cancer, coronary disease, metabolic syndrome, neurodegenerative diseases, and autoimmune diseases [2].

Regulation of Nrf2 on metabolism

Nrf2 improves the gene expression involved in glycolysis, the pentose phosphate pathways, glycogen metabolism, amino acid metabolism, glutaminolysis, fatty acid synthesis, heme metabolism, nucleotide biosynthesis, and glutathione synthesis. Conversely,

Nrf2 decreases the gene expression correlated with gluconeogenesis. Many studies support that Nrf2 is a major key in the reprogramming of cancer metabolism [2]. The regulatory mechanisms of Nrf2 are illustrated in Figure 3.

Nrf2 regulation on unfolded protein response (UPR) and proteostasis

Misfolded protein accumulation triggers the production of ROS from the mitochondria and endoplasmic reticulum (ER), leading to an increase in activation of Nrf2 [17]. Nrf2 activates transcription of ATF4, which is related to amino acid metabolism. In *C. elegans*, the homologous Nrf2 protein SKN-1 activates various UPR target genes, including ATF6 and XBP1, thereby enhancing the UPR program by maintaining ER integrity and protein homeostasis (proteostasis) [21]. Additionally, Nrf2 promotes the expression of proteasome maturation proteins (POMP), which is crucial for proteostasis [22]. In summary, Nrf2 increases proteosome activity and antioxidants expression, aiding in stress adaptation.

Nrf2 regulation on autophagy

Nrf2 plays a significant role in autophagy by upregulating the expression of autophagy-related genes, including SQSTM1/P62, autophagy protein 5 (ATG5), calcium-binding and coiled-coil main-containing protein 2 (CALCOCO2/NDP52), gamma-aminobutyric acid receptor-associated protein-like 1 (GABARAPL1), and unc-51-like kinase 1 (ULK1) [17]. Interestingly, the activation of Nrf2 can be stimulated by the accumulation of oxidized proteins or organelles due to impaired autophagy. This increase in Nrf2 is mediated by the accumulation of p62, a multifunctional receptor that stabilizes KEAP 1 and Nrf2, under autophagy-deficient conditions. In conclusion, a positive feedback loop by p62 and Nrf2 can regulate numerous cellular functions [23].

NRF2 regulation on mitochondrial biogenesis

Nrf2 is involved in enhancing the production and growth of new mitochondria within cells. This

process is known as mitochondrial biogenesis. Nrf2 stimulates mitochondrial biogenesis by activating nuclear respiratory factor-1 (NRF-1), which is a key regulator of mitochondrial transcription factor A (TFAM) and mitochondrial transcription factor B2 (TFBM2) [24]. TFAM is involved in the transcription and maintenance of mitochondrial DNA, while TFBM2 works alongside TFAM to regulate mitochondrial gene expression. Nrf2 also directly regulates the expression of peroxisome proliferator-activated receptor gamma co-activator 1 alpha (PGC-1 α), a principal regulator of mitochondrial function and biogenesis. The cooperation between Nrf2 and PGC-1 α regulates various aspects of mitochondrial function [24–26].

Conclusion

Nrf2 is a widely expressed transcription factor in mammalian cells, belonging to the Cap'n'collar (CNC) protein family. It features a unique CNC domain that binds to the basic leucine zipper (CNC-bZIP) motif. The Nrf2 protein contains seven functional domains, known as Nrf2-ECH homology domains (Neh1-Neh7). Nrf2 regulation occurs at multiple stages, including stability, transcription, post-transcription, post-translation, and target gene activation. This regulation is highly selective, with only specific genes capable of interacting with Nrf2 and its binding partners to achieve activation. Recent studies have highlighted the pleiotropic nature of Nrf2, demonstrating its roles not only in oxidative stress response and drug detoxification but also in metabolism, heme synthesis, proteostasis, unfolded protein response (UPR), autophagy, and mitochondrial biogenesis.

Acknowledgment

None.

Funding

This review received no external funding.

Declaration of interest

The authors declare no conflict of interest.

Author contributions

DY: Conceptualization, Writing – original draft; SWAJ: Supervision, Writing – review and editing.

Received: August 24, 2022

Revised: April 20, 2024

Accepted: June 30, 2024

Published online: July 1, 2024

References

- He F, Ru X, Wen T. NRF2, a transcription factor for stress response and beyond. *Int J Mol Sci.* 2020;21(13):1-23. <https://doi.org/10.3390/ijms21134777>
- He F, Antonucci L, Karin M. NRF2 as a regulator of cell metabolism and inflammation in cancer. *Carcinogenesis.* 2020;41(4):405-16. <https://doi.org/10.1093/carcin/bga039>
- Hayes JD, Dinkova-Kostova AT, Raghunath A, Sundarraj K, Nagarajan R, Arfuso F, et al. NRF2 regulatory network provides an interface between redox and intermediary metabolism. *Trends Biochem Sci.* 2014;39(4):199-218. <https://doi.org/10.1016/j.tibs.2014.02.002>
- Rushmore TH, Morton MR, Pickett CB. The antioxidant responsive element: Activation by oxidative stress and identification of the DNA consensus sequence required for functional activity. *J Biol Chem.* 1991;266(18):11632-9. [https://doi.org/10.1016/S0021-9258\(18\)99004-6](https://doi.org/10.1016/S0021-9258(18)99004-6)
- Moi P, Chan K, Asunis I, Cao A, Kan YW. Isolation of NF-E2-related factor 2 (Nrf2), a NF-E2-like basic leucine zipper transcriptional activator that binds to the tandem NF-E2/AP1 repeat of the β -globin locus control region. *Proc Natl Acad Sci U S A.* 1994;91(21):9926-30. <https://doi.org/10.1073/pnas.91.21.9926>
- Raghunath A, Sundarraj K, Nagarajan R, Arfuso F, Bian J, Kumar AP, et al. Antioxidant response elements: Discovery, classes, regulation and potential applications. *Redox Biol.* 2018; 17(May): 297-314. <https://doi.org/10.1016/j.redox.2018.05.002>
- Chan K, Lu R, Chang JC, Kan YW. NRF2, a member of the NFE2 family of transcription factors, is not essential for murine erythropoiesis, growth, and development. *Proc Natl Acad Sci U S A.* 1996;93(24):13943-8. <https://doi.org/10.1073/pnas.93.24.13943>
- Itoh K, Chiba T, Takahashi S, Ishii T, Igarashi K, Katoh Y, et al. An Nrf2/small Maf heterodimer mediates the induction of phase II detoxifying enzyme genes through antioxidant response elements. *Biochem Biophys Res Commun.* 1997;236(2):313-22. <https://doi.org/10.1006/bbrc.1997.6943>
- Ma Q. Role of Nrf2 in oxidative stress and toxicity. *Annu Rev Pharmacol Toxicol.* 2013;53(1):401-26. <https://doi.org/10.1146/annurev-pharmtox-011112-140320>
- Suzuki T, Shibata T, Takaya K, Shiraishi K, Kohno T, Kunitoh H, et al. Regulatory Nexus of Synthesis and Degradation Deciphers Cellular Nrf2 Expression Levels. *Mol Cell Biol.* 2013;33(12):2402-12. <https://doi.org/10.1128/MB.00065-13>
- Tonelli C, Chio IIC, Tuveson DA. Transcriptional Regulation by Nrf2. *Antioxidants Redox Signal.* 2018;29(17):1727-45. <https://doi.org/10.1089/ars.2017.7342>
- Taguchi K, Fujikawa N, Komatsu M, Ishii T, Unno M, Akaike T, et al. Keap1 degradation by autophagy for the maintenance of redox homeostasis. *Proc Natl Acad Sci U S A.* 2012;109(34):13561-6. <https://doi.org/10.1073/pnas.1121572109>
- Baird L, Yamamoto M. The molecular mechanisms regulating the KEAP1-NRF2 pathway. *Mol Cell Biol.* 2020; 40(13):1-23. <https://doi.org/10.1128/MB.00099-20>
- Ikehata H, Yamamoto M. Roles of the KEAP1-NRF2 system in mammalian skin exposed to UV radiation. *Toxicol Appl Pharmacol.* 2018;360(July):69-77. <https://doi.org/10.1016/j.taap.2018.09.038>
- He F, Antonucci L, Yamachika S, Zhang Z, Taniguchi K, Umemura A, et al. NRF2 activates growth factor genes and downstream AKT signaling to induce mouse and human hepatomegaly. *J Hepatol.* 2020;72:1182-1195. <https://doi.org/10.1016/j.jhep.2020.01.023>
- Mitsui Y, Taguchi K, Kawatani Y, Shibata T, Nukiwa T, Aburanti H, et al. Nrf2 Redirects Glucose and Glutamine into Anabolic Pathways in Metabolic Reprogramming. *Cancer Cell.* 2012;22(1):66-79. <https://doi.org/10.1016/j.ccr.2012.05.016>
- Canning P, Sorrell FJ, Bullock AN. Structural basis of Keap1 interactions with Nrf2. *Free Radic Biol Med.* 2015;88(Part B):101-7. <https://doi.org/10.1016/j.freeradbiomed.2015.05.034>
- Hentze MW, Castello A, Schwarzl T, Preiss T. A brave new world of RNA-binding proteins. *Nat Rev Mol Cell Biol.* 2018;19(5):327-41. <https://doi.org/10.1038/nrm.2017.130>
- Wu KC, Cui JV, Klaassen CD. Effect of graded nrf2 activation on phase-i and -ii drug metabolizing enzymes and transporters in mouse liver. *PLoS One.* 2012;7(7). <https://doi.org/10.1371/journal.pone.0039006>
- Fu J, Xiong Z, Huang C, Li J, Yang W, Han Y, et al. Hyperactivity of the transcription factor Nrf2 causes metabolic reprogramming in mouse esophagus. *J Biol Chem.* 2019;294(1):327-40. <https://doi.org/10.1074/jbc.RA118.005963>
- Jang J, Wang Y, Kim HS, Lalli MA, Kosik KS. Nrf2, a regulator of the proteasome, controls self-renewal and pluripotency in human embryonic stem cells. *Stem Cells.* 2014;32(10):2616-25. <https://doi.org/10.1002/stem.1764>
- Pajares M, Cuadrado A, Rojo AI. Modulation of proteostasis by transcription factor NRF2 and impact in neurodegenerative diseases. *Redox Biol.* 2017;11(January):543-53. <https://doi.org/10.1016/j.redox.2017.01.006>

23. Uruno A, Furusawa Y, Yagishita Y, Fukutomi T, Muramatsu H, Negishi T, et al. The Keap1-Nrf2 System Prevents Onset of Diabetes Mellitus. *Mol Cell Biol*. 2013;33(15):2996-3010. <https://doi.org/10.1128/MCB.00225-13>
24. Gureev AP, Shaforostova EA, Popov VN. Regulation of mitochondrial biogenesis as a way for active longevity: Interaction between the Nrf2 and PGC-1 α signaling pathways. *Front Genet*. 2019; 10(May): 1-12. <https://doi.org/10.3389/fgene.2019.00435>
25. Piantadosi CA, Carraway MS, Babiker A, Suliman HB. Heme oxygenase-1 regulates cardiac mitochondrial biogenesis via nrf2-mediated transcriptional control of nuclear respiratory factor-1. *Circ Res*. 2008;103(11):1232-40. <https://doi.org/10.1161/01.RES.0000338597.71702.ad>
26. Hu Q, Ren J, Li G, Wu J, Wu X, Wang G, et al. The mitochondrially targeted antioxidant MitoQ protects the intestinal barrier by ameliorating mitochondrial DNA damage via the Nrf2/ARE signaling pathway. *Cell Death Dis*. 2018;9(3). <https://doi.org/10.1038/s41419-018-0436-x>

The role of Nrf2 transcription factors in various physiological and pathological states

ORIGINALITY REPORT

19%
SIMILARITY INDEX 12%
INTERNET SOURCES 21%
PUBLICATIONS 7%
STUDENT PAPERS

PRIMARY SOURCES

1	Systems Biology of Free Radicals and Antioxidants, 2014. Publication	1 %
2	Walter H. Moos, Douglas V. Faller, Ioannis P. Glavas, David N. Harpp et al. "Epigenetic treatment of dermatologic disorders", Drug Development Research, 2019 Publication	1 %
3	Submitted to University of Dundee Student Paper	1 %
4	Danyelly Bruneska Gondim Martins, Thaysa Walleria Aragão Santos, Maria Helena Menezes Estevam Alves, Rosângela Ferreira Frade de Araújo et al. "The Role of NRF2 Transcription Factor in Metabolic Syndrome", IntechOpen, 2024 Publication	1 %
5	encyclopedia.pub Internet Source	1 %
6	repositorio.uam.es Internet Source	1 %
7	journals.innovareacademics.in Internet Source	1 %
8	research.aston.ac.uk Internet Source	1 %
9	Yongjie Cen, Xiaopeng Zou, Qien Zhong, Yumei Chen, Yiguang Lin, Qili Feng, Xiaoyun	1 %

Wang, Sichun Zheng. "The TIAR-mediated Nrf2 response to oxidative stress is mediated through the Nrf2 noncoding 3'untranslated region in *Spodoptera litura*", *Free Radical Biology and Medicine*, 2022

Publication

10 Marc Pfefferlé, Florence Vallelian. "Chapter 8 Transcription Factor NRF2 in Shaping Myeloid Cell Differentiation and Function", Springer Science and Business Media LLC, 2024 1 %

Publication

11 link.springer.com 1 %

Internet Source

12 www.preprints.org 1 %

Internet Source

13 Submitted to University of Birmingham 1 %

Student Paper

14 Submitted to University of Nevada, Las Vegas 1 %

Student Paper

15 f.oads.cc 1 %

Internet Source

16 Lei Liu, Logan M. Locascio, Sylvain Doré. "Critical Role of Nrf2 in Experimental Ischemic Stroke", *Frontiers in Pharmacology*, 2019 1 %

Publication

17 Submitted to Victoria University of Wellington 1 %

Student Paper

18 Zhu, Jiayu, Huihui Wang, Feng Chen, Jingqi Fu, Yuanyuan Xu, Yongyong Hou, Henry H. Kou, Cheng Zhai, M. Bud Nelson, Qiang Zhang, Melvin E. Andersen, and Jingbo Pi. "An overview of chemical inhibitors of the Nrf2-ARE signaling pathway and their potential 1 %

applications in cancer therapy", Free Radical Biology and Medicine, 2016.

Publication

19 Claudia Tonelli, Iok In Christine Chio, David A. Tuveson. "Transcriptional Regulation by Nrf2", Antioxidants & Redox Signaling, 2018 1 %
Publication

20 Tian Wang, Xi-Ya Sun, Ai-Ling Li, Ming-Xing Zhou et al. "Botrysphin D attenuates arsenic-induced oxidative stress in human lung epithelial cells via activating Nrf2/ARE signaling pathways", Biochemical and Biophysical Research Communications, 2019 1 %
Publication

21 www.frontiersin.org 1 %
Internet Source

22 www.spandidos-publications.com 1 %
Internet Source

23 Zhiguo Zhang, Shanshan Zhou, Xin Jiang, Yue-Hui Wang, Fengsheng Li, Yong-Gang Wang, Yang Zheng, Lu Cai. "The role of the Nrf2/Keap1 pathway in obesity and metabolic syndrome", Reviews in Endocrine and Metabolic Disorders, 2014 <1 %
Publication

24 e-crt.org <1 %
Internet Source

25 www.imrpress.com <1 %
Internet Source

26 www.liebertpub.com <1 %
Internet Source

27 www.wcrj.net <1 %
Internet Source

Exclude quotes Off

Exclude bibliography On

Exclude matches < 10 words

The role of Nrf2 transcription factors in various physiological and pathological states

GRADEMARK REPORT

FINAL GRADE

GENERAL COMMENTS

/0

PAGE 1

PAGE 2

PAGE 3

PAGE 4

PAGE 5

PAGE 6

PAGE 7
