

Subchronic kretek smoke exposure induces myocardial p53 protein expression and associated inflammatory and fibrotic alterations in rats

David Tjahyadi, Hosea Jaya Edy¹,
Endrico Xavierees²,
Joey Joshua Vidova Tjahyadi³,
Laurentia Gabrielle³,
Mellysa Henggar Ratnasari³,
Marlita Sabta Utari³,
Ashaolu Victoria Oladimeji⁴,
Seçil Karahüseyin⁵

Department of Histology, Faculty of Medicine, Universitas Trisakti, Jakarta,
¹Pharmacy Study Program, Faculty of Mathematics and Natural Sciences, Universitas Sam Ratulangi, Manado,
²Department of Biochemistry, Faculty of Medicine, Universitas Trisakti,
³Medical Education Program, Faculty of Medicine, Universitas Trisakti, Jakarta, Indonesia, ⁴Department of Chemistry, Loyola Institute of Frontier Energy, Loyola College, University of Madras, Chennai, Tamil Nadu, India,
⁵Department of Pharmacognosy, Faculty of Pharmacy, Cukurova University, Adana, Turkey

J. Adv. Pharm. Technol. Res.

ABSTRACT

Kretek smoke contains various toxic substances that are harmful to human health and has been associated with inflammation and structural alterations in multiple organs. However, its effects on cardiac tissue remain incompletely understood. This study aimed to evaluate myocardial histopathological alterations and p53 protein expression following subchronic kretek smoke exposure in rats. Male *Rattus norvegicus* Sprague–Dawley rats were randomly divided into two groups. Group I was exposed to normal air, while Group II was exposed to kretek smoke at a dose of 1 kretek/rat/day for 30 days. Cardiac tissues were examined using H&E and MT staining. IHC analysis was performed to assess myocardial p53 protein expression, while ultrastructural observations of the myocardium were conducted using SEM. Group II rats exhibited myocardial inflammation characterized by lymphocyte infiltration and increased collagen accumulation, whereas Group I rats showed no inflammatory changes and minimal collagen deposition. Myocardial p53 protein expression was significantly higher in Group II ($224.68 \pm 5.44 \mu\text{m}^2$) than in Group I ($199.87 \pm 13.35 \mu\text{m}^2$; $P = 0.005$). Ultrastructural myocardial alterations were observed in Group II but not in Group I. In conclusion, subchronic kretek smoke exposure increases myocardial p53 protein expression accompanied by inflammatory and fibrotic alterations.

Key words: Fibrosis, inflammation, kretek smoke exposure, myocardium, p53 protein expression

INTRODUCTION

Most smokers in Indonesia use clove-mixed tobacco cigarettes known as kreteks.^[1] This condition increases the number of individuals exposed to secondhand smoke.^[2]

Address for correspondence:

Dr. David Tjahyadi,
Department of Histology, Faculty of Medicine,
Universitas Trisakti, Jakarta, Indonesia.
E-mail: davesaboch@trisakti.ac.id

Submitted: 18-Jan-2026

Revised: 10-Apr-2026

Accepted: 22-Apr-2026

Published: 12-Aug-2026

Access this article online

Quick Response Code:



Website:

<https://www.ovid.com/jnls/japtr>

DOI:

10.4103/JAPTR.JAPTR_12_26

Both active smokers and passive smokers remain at risk of smoking-related diseases.^[3] Smoking prevalence in Indonesia remains high and represents an important public health concern in the ASEAN region.^[4] Tobacco exposure also contributes to cancer burden and premature mortality, including among individuals exposed to secondhand smoke.^[5] Smoking is also associated with oral mucosal lesions such as leukoplakia in exposed individuals.^[6] These conditions highlight the importance of investigating smoke-related injury in the cardiovascular system.

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 License (CC BY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

For reprints contact: WKHLRPMedknow_reprints@wolterskluwer.com

How to cite this article: Tjahyadi D, Edy HJ, Xavierees E, Tjahyadi JJ, Gabrielle L, Ratnasari MH, *et al.* Subchronic kretek smoke exposure induces myocardial p53 protein expression and associated inflammatory and fibrotic alterations in rats. *J Adv Pharm Technol Res* 2026;17:288-95.

Cigarette smoke exposure induces cellular signaling alterations in airway smooth muscle cells through TRPA1-mediated calcium influx.^[7] Cigarette smoke combustion also produces particulate emissions that contribute to biological exposure during inhalation.^[8] Smoke particles deposit in the respiratory tract under realistic inhalation conditions and contribute to tissue exposure mechanisms.^[9] Previous studies demonstrated decreased density of pyramidal cells in the cerebral cortex and Purkinje cells in the cerebellum after exposure to filtered kretek cigarette smoke in rats.^[10] Additional studies also reported histometric abnormalities of Purkinje cells following low-dose kretek smoke exposure.^[11] These findings indicate that kretek smoke exposure produces measurable structural alterations in neural tissue.

The heart is also vulnerable to cigarette smoke exposure because smoke-derived toxic compounds can reach systemic circulation and contribute to myocardial injury processes. Experimental studies demonstrated that cigarette smoke exposure induces cardiac hypertrophy and vascular inflammation in animal models.^[12] Short-term cigarette smoke exposure also induces cardiac remodeling in rats.^[13] Smoke exposure is further associated with inflammatory mechanisms that contribute to tissue injury processes.^[14] Our previous study demonstrated increased p53 gene expression in lung tissue after exposure to low doses of kretek cigarette smoke in rats.^[15]

Evidence linking kretek smoke exposure with myocardial histopathological alteration remains limited. Information regarding myocardial p53 protein expression after kretek smoke exposure is also still scarce. Therefore, this study evaluated myocardial histopathological alterations in rats exposed to subchronic kretek smoke. The study also assessed collagen accumulation in myocardial tissue. In addition, p53 protein immunopositivity was examined in exposed rats.

MATERIALS AND METHODS

Type of research

This randomized controlled study used male *Rattus norvegicus* rats of the Sprague–Dawley strain. Animals were randomly assigned to two groups, Group I (control) and Group II (kretek smoke exposure).

Nicotine content in kretek

Nicotine content in kretek was measured using high-performance thin-layer chromatography. A CAMAG Linomat 5 instrument (CAMAG, Switzerland; Linomat 5_220727, S/N 220727, version 1.00.13) was used. Kretek samples were extracted and analyzed in triplicate. Nicotine concentration was expressed as mean $\pm$ standard deviation (SD) ($\mu\text{g/g}$) to ensure accuracy and reproducibility.

Sample size and animals

Sample size was guided by the resource equation, with five animals per group considered sufficient for exploratory studies.^[16] Similar designs using Sprague–Dawley rats for histological and p53 analyses have been reported previously.^[11,15] All rats were male, aged 2–3 months, and weighed 200–250 g. Rats were anesthetized with ketamine 100 mg/kg and xylazine 10 mg/kg, intraperitoneal, prior to euthanasia, as previously described.^[10] Hearts were collected and fixed in 10% neutral buffered formalin (NBF) for histological preparation and p53 immunohistochemical analysis.

Animal treatment

Rats were housed individually with *ad libitum* food and water. Group I breathed normal air, and Group II was exposed to 1 kretek/rat/day for 30 days in a custom chamber. At the end of exposure, rats were anesthetized, and hearts were collected and fixed in NBF. A schematic of the study design is shown in Figure 1.

Histological analysis

Heart tissues were sectioned at 5 μm . H and E staining evaluated myocardial histomorphology and inflammation, while Masson's Trichrome (MT) staining assessed collagen deposition. Observations were performed at $\times 400$ by three independent observers blinded to group allocation. Five random fields per section were captured per animal. Collagen area fraction was quantified using ImageJ with threshold-based segmentation. The mean per animal was used as the biological replicate for analysis.

Scanning electron microscopy

Tissue samples were coated with platinum using a Turbo Pumped Sputter Coater Q150T (Quorum Technologies, United Kingdom). Ultrastructural analysis was performed using a FEI Nova NanoSEM 450 (Thermo Fisher Scientific, Czech Republic) at accelerating voltages of 2–5 kV. Scanning electron microscopy (SEM) was used to evaluate ultrastructural myocardial morphology.

Immunohistochemistry for p53 protein

p53 protein expression was evaluated by immunohistochemistry. Deparaffinized sections underwent antigen retrieval in citrate buffer (pH 6.0) at 95°C for 20 min, followed by incubation with anti-p53 primary antibody (mouse monoclonal, clone DO-7, NCL-L-p53, Leica Biosystems; 1:550). horseradish peroxidase-conjugated secondary antibody and 3,3'-diaminobenzidine chromogen were applied for 10 min at 25°C. Sections were counterstained with hematoxylin. Brown nuclear staining indicated positive p53 expression. Quantification was performed in five random fields per section using ImageJ with color deconvolution; the mean per animal was used as a biological replicate. Observers were blinded to group allocation. Results are expressed as mean $\pm$ SD.

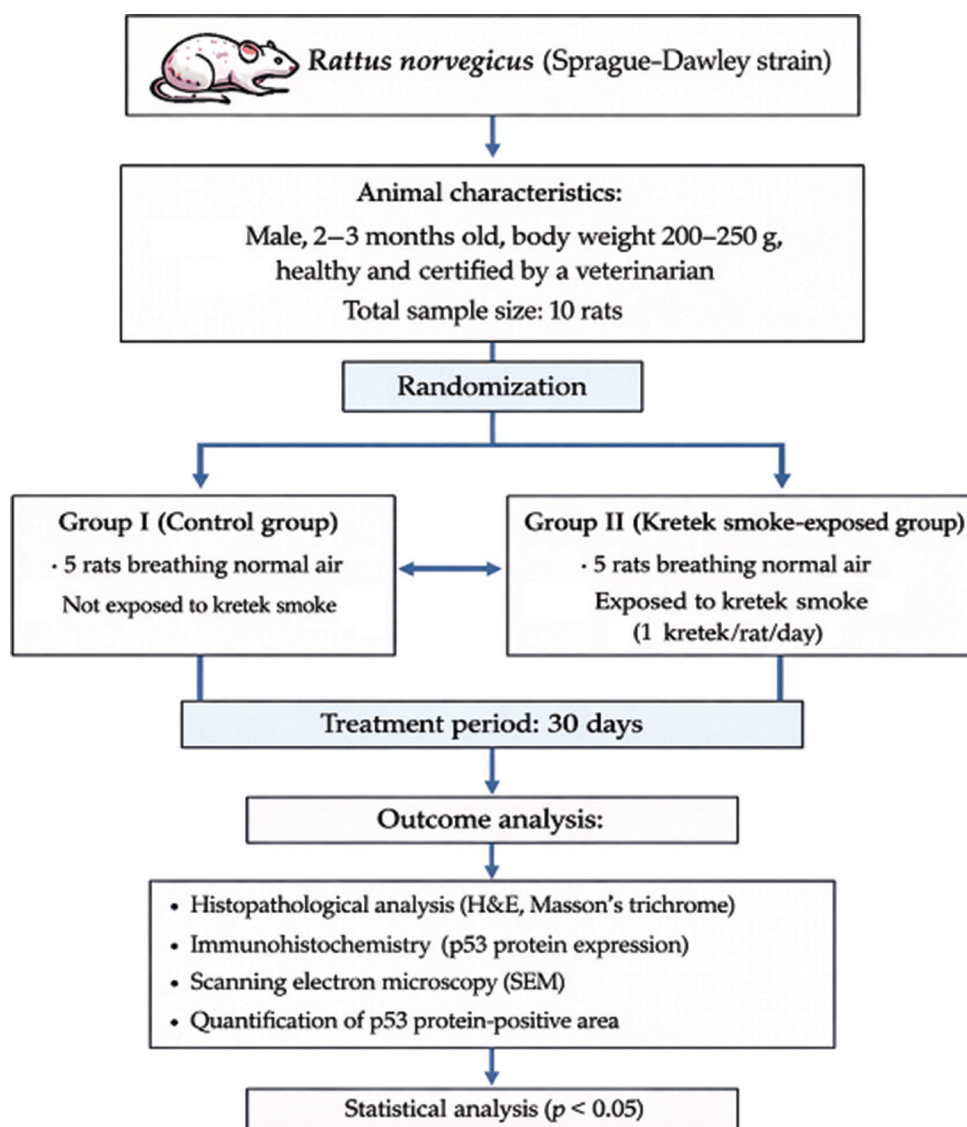


Figure 1: Experimental design of kretek smoke exposure in rats. Group I (control group) rats breathed normal air. Group II (kretek smoke-exposed group) rats received smoke exposure (1 kretek/rat/day) for 30 days. H and E: Hematoxylin-eosin

Statistical analysis

Five random fields per heart section were analyzed for collagen deposition. The mean collagen area fraction per animal was used as the unit of analysis ($n = 5$ per group). Data normality was assessed using the Shapiro–Wilk test and homogeneity of variances using Levene's test. Because assumptions were met, independent t -tests were used to compare collagen area fraction and p53 expression between groups. $P < 0.05$ was considered statistically significant.

Ethical approval

This study was approved by the Research Ethics Committee, Faculty of Medicine, Universitas Trisakti (No. 011/KER/FK/03/2025). All procedures followed internationally accepted guidelines for laboratory animal care.

RESULTS

Heart morphology

Gross examination showed normal cardiac shape and uniform dark red coloration in both groups, with no visible surface abnormalities. The average heart length was 3.1 cm, and the average width was 2.3 cm, with no apparent differences between Group I and Group II [Figure 2].

Nicotine content in kretek

Nicotine concentration in kretek extract was 3.88 ± 0.05 $\mu\text{g/g}$.

Histological features of the rat myocardial tissue

Histological examination of Group I is shown in Figure 3a-c. The myocardial tissue exhibited normal structure, with preserved endocardium, myocardium, epicardium, and pericardium. No vascular inflammation or lymphocyte infiltration was

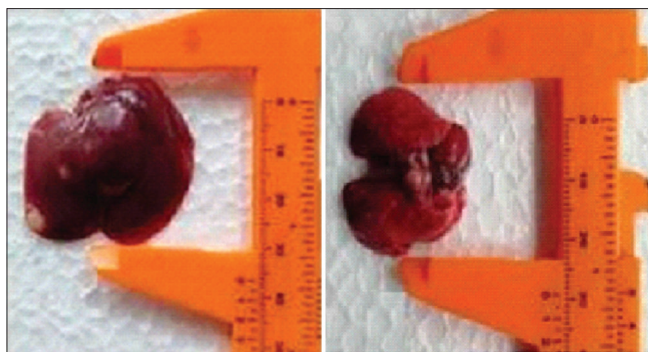


Figure 2: Gross morphology of the heart from *Rattus norvegicus* (Sprague-Dawley strain) rats

observed (blue arrows). In contrast, Group II rats showed structural alterations [Figure 3d-f], including vascular inflammation and lymphocyte infiltration (red arrows). The severity and extent of lesions varied among animals.

Scanning electron microscopy of rat myocardium

SEM images [Figure 4] revealed morphological differences in myocardial fibers between Group I (control) and Group II (smoke-exposed) rats. Because SEM primarily shows surface morphology, qualitative sarcomere enlargement in Group II should be interpreted with caution, and quantitative assessment requires TEM or fluorescence-based imaging.

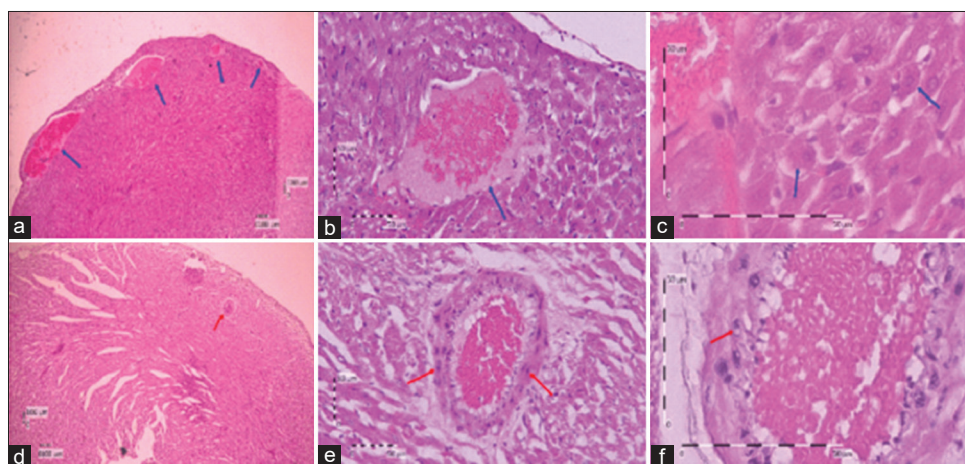


Figure 3: Comparison of rat heart histology between Group I and Group II. (a-c) Heart histology of Group I, (d-f) heart histology of Group II. All sections were stained with hematoxylin-eosin, (a and d) $\times 40$ magnification, (b and e) $\times 400$ magnification, (c and f) $\times 1000$ magnification. The blue arrows indicate the absence of inflammation in cardiac blood vessels, whereas the red arrows indicate inflammation of cardiac blood vessels characterized by lymphocyte infiltration. Group I (control group) rats breathed normal air. Group II (kretek smoke-exposed group) rats received smoke exposure (1 kretek/rat/day) for 30 days

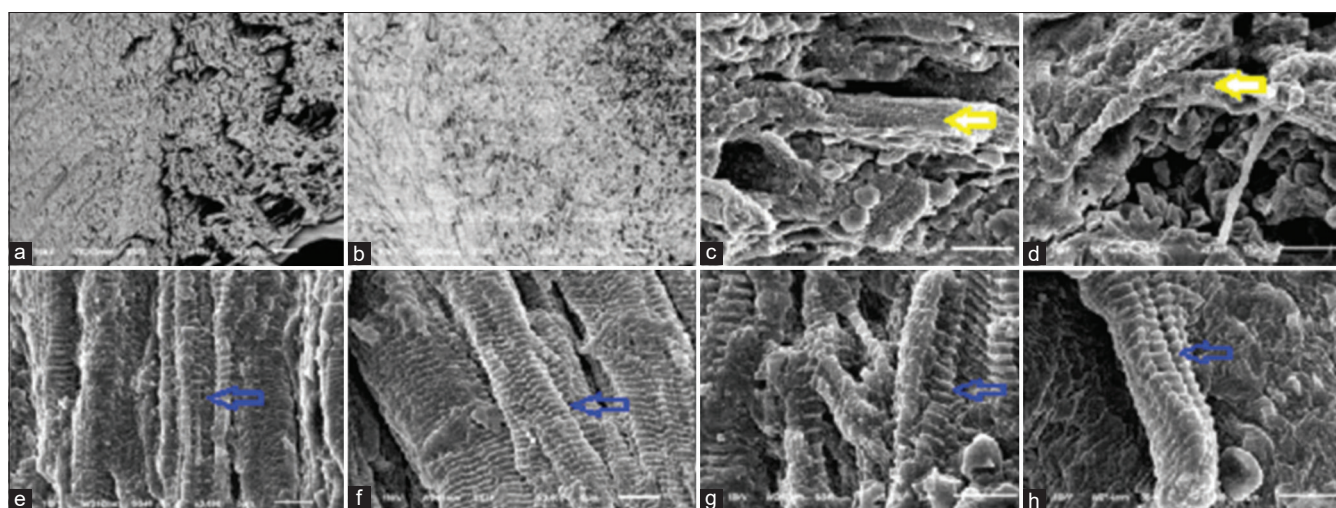


Figure 4: Scanning electron microscopy images of rat cardiac muscle, (a, c, e and g) Group I, (b, d, f, and h) Group II, (a and b) transverse sections at $\times 100$ magnification, (c and d) $\times 2500$ magnification, (e and f) longitudinal sections at $\times 3000$, and (g and h) at $\times 5000$ magnification. Yellow arrows indicate ventricular cardiomyocytes, and blue arrows indicate sarcomeres. Group I (control group) rats breathed normal air. Group II (kretek smoke-exposed group) rats received smoke exposure (1 kretek/rat/day) for 30 days

Myocardial inflammation

Kretek smoke exposure induced vascular inflammation in Group II myocardium. Inflammation areas ranged from 3455.09 to 7030.43 μm^2 , and lesion areas from 1032.87 to 3305.50 μm^2 . Mean inflammation ($5241.57 \pm 1669.60 \mu\text{m}^2$) exceeded mean lesion ($2079.22 \pm 946.82 \mu\text{m}^2$). The coefficient of variation was 31.85% for inflammation and 45.54% for lesion, indicating substantial inter-animal variability. Boxplot and histology [Figure 5a and b] confirmed lymphocyte infiltration; inflammation areas were larger and more variable than lesions [Supplementary Table 1].

Myocardial collagen accumulation

MT staining showed minimal collagen deposition in Group I, whereas Group II exhibited multifocal collagen

accumulation, with a mean collagen area fraction of $21.98\% \pm 0.58\%$ [Figure 6 and Supplementary Table 2]. Collagen deposition in Group I remained minimal and was therefore not subjected to quantitative analysis due to near-baseline staining levels.

p53 protein expression

The region of interest area positive for p53 protein expression was significantly higher in Group II ($224.68 \pm 5.44 \mu\text{m}^2$) than in Group I ($199.87 \pm 13.35 \mu\text{m}^2$) ($t(8) = -3.849$, $P = 0.005$; Figure 7c [Supplementary Table 3]). The mean difference between groups was $24.81 \mu\text{m}^2$ (95% confidence interval [CI]: 9.94–39.67). Bootstrap analysis (1000 resamples) confirmed the robustness of this difference ($P = 0.034$; 95% CI: –38.03 to –13.37).

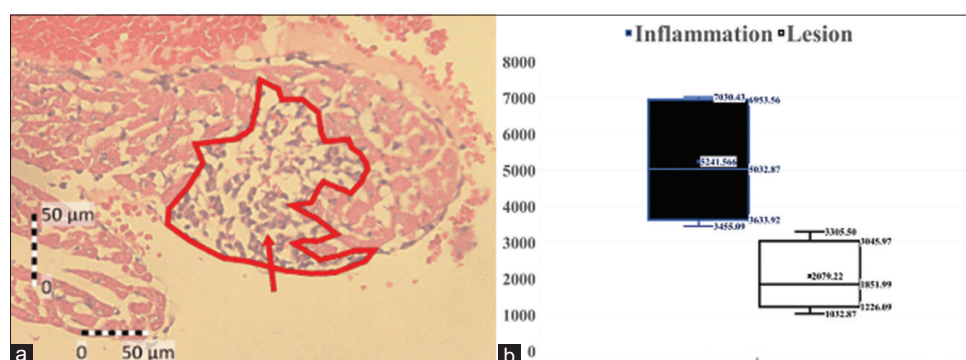


Figure 5: Histological evidence of myocardial inflammation in Group II rats, (a) photomicrograph of myocardial tissue stained with hematoxylin-eosin at $\times 400$ magnification. Red arrows indicate lymphocyte infiltration, showing multifocal inflammatory foci, (C2) (b) Boxplot showing the distribution of inflammatory and lesion areas in Group II myocardium. Each point represents the mean area from five randomly selected fields per animal. Group II (kretek smoke-exposed group) rats received smoke exposure (1 kretek/rat/day) for 30 days

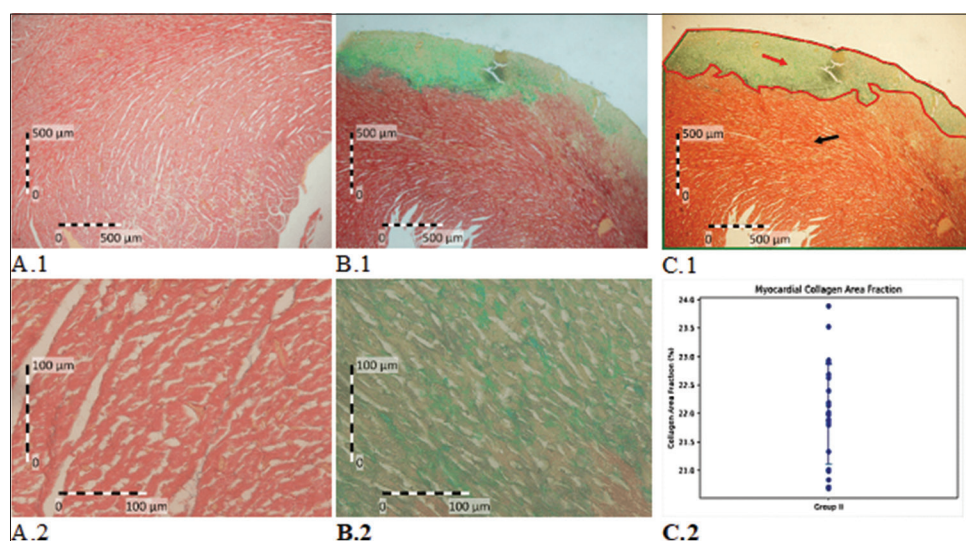


Figure 6: Myocardial collagen assessment, (A1 and A2) histology of Group I with Masson's Trichrome ($\times 40$, $\times 400$), (B1 and B2) histology of Group II under same conditions, (C1) region of interest for histomorphometry in Group II ($\times 40$), collagen in green (red arrows), muscle fibers in red (black arrows), (C2) Scatter plot of myocardial collagen fraction in Group II ($n = 5$), each dot = mean of five fields per animal, central marker = mean, error bars = standard deviation. Group I (control group) rats breathed normal air. Group II (kretek smoke-exposed group) rats received smoke exposure (1 kretek/rat/day) for 30 days. A1 = Group I ($\times 40$), A2 = Group I ($\times 400$), B1 = Group II ($\times 40$), B2 = Group II ($\times 400$), C1 = Region of Interest for Histomorphometry in Group II, C2 = Scatter Plot of Myocardial Collagen Area Fraction in Group II

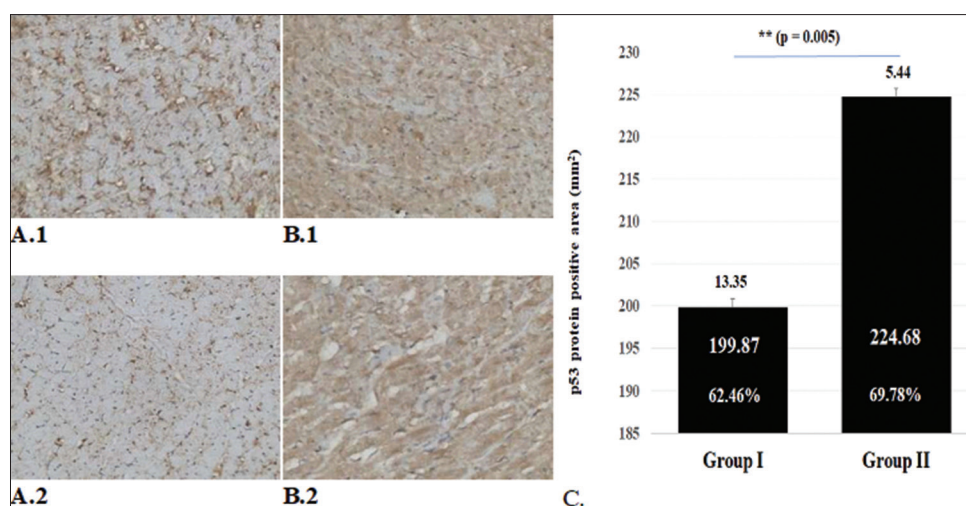


Figure 7: p53 protein expression in rat cardiac muscle detected by immunohistochemistry (IHC) (A1 and A2) IHC images of cardiac muscle from Group I, (B1 and B2) IHC images from Group II. All sections were examined at $\times 200$ magnification, and (C) the analysis of p53 expression. Group I (control group) rats breathed normal air. Group II (kretek smoke-exposed group) rats received smoke exposure (1 kretek/rat/day) for 30 days. A1 = Group I (IHC image), A2 = Group I (IHC image), B1 = Group II (IHC image), B2 = Group II (IHC image), C = Quantitative Analysis of p53 Protein Expression

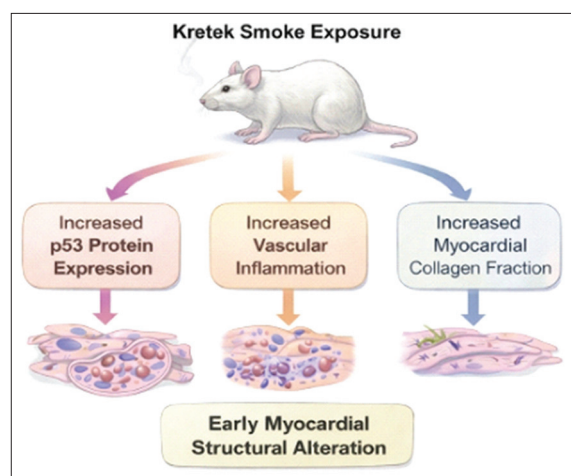


Figure 8: Schematic illustration of myocardial alterations following subchronic kretek smoke exposure in rats. Subchronic exposure to kretek cigarette smoke (1 cigarette/rat/day for 30 days) was associated with increased myocardial p53 protein expression, vascular inflammation characterized by inflammatory cell infiltration, and increased myocardial collagen area fraction in exposed rats (Group II). These findings indicate early myocardial structural alterations following kretek smoke exposure

DISCUSSION

In this study, rats exposed to kretek smoke for 30 days experienced myocardial inflammation, structural alterations, and collagen accumulation. These findings were accompanied by increased nuclear p53 expression and qualitative sarcomere enlargement. Therefore, subchronic kretek smoke exposure induces structural and molecular alterations in myocardial tissue, representing early alterations in the pathogenesis of cardiovascular disease.

This study demonstrated myocardial abnormalities in rats. This is in line with previous research showing that subchronic smoke exposure induces myocardial inflammation,^[13] direct cardiomyocyte toxicity and mitochondrial dysfunction,^[17] and multifocal collagen deposition.^[18] These findings are consistent with smoke-induced cardiac remodeling reported in experimental models.^[13] Furthermore, it contributes to the observed collagen accumulation. Structural alterations accompanied by increased nuclear p53 protein expression are also in line with studies demonstrating stress responses in cardiomyocytes.^[19] Sarcomere enlargement in this study reflects ultrastructural adaptation to chronic injury, as in previous studies.^[20] Collectively, our findings indicate that subchronic kretek smoke exposure induces structural and molecular alterations in myocardial tissue. These results are consistent with mechanisms of myocardial fibrosis and early remodeling in cardiovascular injury.^[21]

In this study, lymphocyte infiltration and vascular inflammation were observed in Group II. These findings are consistent with previous studies demonstrating that cigarette smoke induces oxidative stress and systemic inflammation, contributing to endothelial dysfunction and cardiovascular risk.^[22] Similar inflammatory responses have been reported in experimental models of cigarette smoke exposure.^[13] Cigarette smoke generates reactive oxygen species (ROS) and free radicals that damage cellular membranes, proteins, and deoxyribonucleic acid (DNA), leading to cardiomyocyte injury.^[23] Cigarette smoke promotes ferroptosis and inflammation in experimental models.^[24] Oxidative stress is enhanced by increased circulating lipopolysaccharides after tobacco smoke exposure.^[25] Clinical evidence also shows increased oxidative stress biomarkers and inflammatory

mediators in smokers, reflecting vascular dysfunction and systemic injury.^[23]

Masson's trichrome staining revealed minimal collagen in rats from Group I. In contrast, rats from Group II showed increased collagen deposition, a hallmark of cardiac fibrosis. These findings are consistent with previous studies demonstrating extracellular matrix remodeling that impairs myocardial compliance and contributes to functional deterioration.^[26] Increased collagen accumulation in this study is consistent with findings from other smoke exposure models.^[27] Previous studies demonstrate that cigarette smoke alters lipid profiles, increases cardiovascular risk, and aggravates myocardial injury.^[28] Sarcomere enlargement in this study represents an adaptive response to increased mechanical load under fibrotic conditions.^[20] Furthermore, biomechanical deterioration in hypertrophied myocardium under chronic stress confirms adverse cardiac remodeling.^[29] Finally, these structural adaptations observed in this study predict poor cardiac outcomes in fibrotic heart disease, consistent with previous research findings.^[30]

The results of this study demonstrate increased p53 protein expression, consistent with previous studies on cellular oxidative stress induced by cigarette smoke exposure.^[19] Previous studies demonstrate that oxidative stress and DNA damage regulate apoptotic and senescence pathways in cardiomyocytes.^[31] ROS-mediated DNA damage directly induces p53 signaling pathways in cardiomyocytes.^[32] Previous studies also demonstrate that p53 activation contributes to inflammatory and fibrotic responses in myocardial tissue.^[31] However, this study evaluated p53 protein expression only. Functional modulation of p53 signaling was not performed. Therefore, further studies are required to clarify the causal role of p53 activation in cigarette smoke-induced myocardial injury.

In this study, sarcomere enlargement was observed, suggesting structural remodeling under chronic stress conditions. These findings are consistent with previous studies demonstrating adaptive and ultrastructural alterations of sarcomere organization during myocardial remodeling under increased mechanical load in fibrotic myocardium.^[29] Such structural alterations are associated with impaired myocardial biomechanics and contractility and are consistent with collagen accumulation observed in this study.^[26]

In this study, subchronic kretek smoke exposure induced heart inflammation and structural alterations. SEM analysis of the myocardium revealed alterations in the ultrastructural organization of cardiomyocytes and sarcomeres. Histological analysis showed collagen accumulation and increased nuclear p53 protein expression. These findings support previous evidence that short-term smoke exposure can damage the heart.^[13] Tobacco smoke is

linked to cardiovascular disease, including ischemic heart disease, and worsens clinical outcomes.^[33] Antioxidants such as ascorbic acid reduce smoke-induced oxidative stress and heart and lung damage in rats.^[34] Early prevention is essential. Public health measures, including smoking cessation and smoke-free environments, are important to protect cardiovascular health.

The overall myocardial alterations observed after 30 days of kretek smoke exposure, including increased p53 protein expression, vascular inflammation, and increased myocardial collagen area fraction, suggest early myocardial structural alteration in exposed rats. These integrated findings are summarized schematically in Figure 8.

Limitations

This study did not include functional cardiac assessments, such as echocardiography or hemodynamic measurements, limiting the interpretation of physiological impact. Ultrastructural observations were qualitative, without precise quantitative metrics. While p53 expression increased, the study design cannot establish causality between p53 activation and myocardial alterations.

CONCLUSION

Subchronic kretek smoke exposure (1 kretek/rat/day for 30 days) induced myocardial inflammation, structural alterations, and increased collagen deposition. Nuclear p53 protein expression was significantly higher in the Group II ($224.68 \pm 5.44 \mu\text{m}^2$) compared to the Group I ($199.87 \pm 13.35 \mu\text{m}^2$; $P = 0.005$), indicating increased cellular stress responses. SEM also revealed ultrastructural alterations of cardiac muscle fibers, supporting the presence of inflammatory and fibrotic myocardial alterations.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

1. Braun M, Marsidi LL, Klingelhöfer D, *et al.* Kretek cigarettes and particulate matter emissions—an aerosol spectrometric study on typical Indonesian brands flavored with cloves. *Nicotine Tob Res* 2022;24:778-84. <https://doi.org/10.1093/ntr/ntab209>.
2. World Health Organization (WHO). Tobacco; 2020. Available from: <https://www.who.int/news-room/fact-sheets/detail/tobacco>.
3. Jha P. The hazards of smoking and the benefits of cessation: A critical summation of the epidemiological evidence in high-income countries. *Elife* 2020;9:e49979.
4. Southeast Asia Tobacco Control Alliance (SEATCA). Tobacco Control Atlas ASEAN Region. 4th ed. Bangkok, Thailand: Southeast Asia Tobacco Control Alliance (SEATCA); 2018. Available from: https://seatca.org/dmdocuments/TobaccoControlAtlas_ASEANRegion_4thEd_Dec2019.pdf. [Last accessed on 2025 Mar 21].

5. Kristina SA, Wiedyaningsih C, Masrida WO, Santoso KA, Ahsan A. The mortality costs of tobacco related cancers among secondhand smokers in Indonesia 2018. *Res J Pharm Technol* 2019;12:6075-80. doi:10.5958/0974-360X.2019.01055.2.
6. Ravichandiran R, Venkatesh J, Felix AJW, Ramya K. Prevalence of oral mucosal lesion in patients with tobacco related habits among Chidambaram population: A cross-sectional study. *Res J Pharm Technol* 2022;15:3377-81. doi:10.52711/0974-360X.2022.00565.
7. Lin J, Taggart M, Borthwick L, Fisher A, Brodlie M, Sassano MF, *et al.* Acute cigarette smoke or extract exposure rapidly activates TRPA1-mediated calcium influx in primary human airway smooth muscle cells. *Sci Rep* 2021;11:9643.
8. Amorós-Pérez A, Cano-Casanova L, Román-Martínez MC, Lillo-Ródenas MÁ. Comparison of particulate matter emission and soluble matter collected from combustion cigarettes and heated tobacco products using a setup designed to simulate puffing regimes. *Chem Eng J Adv* 2021;8:100144. doi:10.1016/j.cej.2021.100144.
9. Paul AR, Khan F, Jain A, Saha SC. Deposition of smoke particles in human airways with realistic waveform. *Atmosphere* 2021;12:912. doi:10.3390/atmos12070912.
10. Tjahyadi D, Parwanto E, Amalia H, Digambiro RA, Edy HJ, Oladimeji AV. Decreased density of pyramidal cells in the cerebral cortex and Purkinje cells in the cerebellar cortex of Sprague-Dawley rats after being exposed to filtered kretek cigarette smoke. *J Biol Res* 2023;96:10757. doi:10.4081/jbr.2023.10757.
11. Tjahyadi D, Parwanto E, Edy HJ, Tjahyadi JJV, Gabrielle L, Oladimeji AV, *et al.* Histometric analysis to perform Purkinje cell abnormalities in rats due to low doses of kretek smoke exposure. *Res J Pharm Technol* 2025;18:4882-90. doi: 10.52711/0974-360X.2025.00704.
12. Al Hariri M, Zibara K, Farhat W, Hashem Y, Soudani N, Al Ibrahim F, *et al.* Cigarette smoking-induced cardiac hypertrophy, vascular inflammation and injury are attenuated by antioxidant supplementation in an animal model. *Front Pharmacol* 2016;7:397.
13. Bocalini DS, Luiz RS, Silva KA, Serra AJ, Avila RA, Leopoldo AS, *et al.* Short-term cigarette smoking in rats impairs physical capacity and induces cardiac remodeling. *Biomed Res Int* 2020;2020:2589892. doi:10.1155/2020/2589892.
14. Bowman WS, Schmidt RJ, Sanghar GK, Thompson III GR, Ji H, Zeki AA, *et al.* Wildfire smoke-induced mechanisms of airway inflammation. *Int Arch Allergy Immunol* 2024;185:600-16. [doi: 10.1159/000536578].
15. Parwanto E, Tjahyadi D, Sisca S, Amalia H, Hairunisa N, Edy HJ, *et al.* Low Doses of Kretek Cigarette Smoke Altered Rat Lung Histometric, and Overexpression of the p53 Gene. *Open Respir Med J* 2024;18:e18743064285619.
16. Arifin WN, Zahiruddin WM. Sample size calculation in animal studies using resource equation approach. *Malays J Med Sci* 2017;24:101-5.
17. Matsumura S, Yasuda J, Notomi T, Suzuki Y, Chen IS, Murakami D, *et al.* Direct toxicity of cigarette smoke extract on cardiac function mediated by mitochondrial dysfunction in Sprague-Dawley rat ventricular myocytes and human induced pluripotent stem cell-derived cardiomyocytes. *PLoS One* 2024;19:e0295737.
18. Aid J, Tanjeko AT, Serré J, Eggelbusch M, Noort W, de Wit GM, *et al.* Smoking cessation only partially reverses cardiac metabolic and structural remodeling in mice. *Acta Physiol (Oxf)* 2024;240:e14145.
19. Jarabíková I, Horváth C, Hrdlička J, Boroš A, Olejníčková V, Záborská E, *et al.* Necrosis like cell death modes in heart failure: The influence of aetiology and the effects of RIP3 inhibition. *Basic Res Cardiol* 2025;120:373-92. [doi: 10.1007/s00395-025-01101-4].
20. Al Hariri M, Zibara K, Farhat W, Hashem Y, Soudani N, Al Ibrahim F, *et al.* Cigarette smoking-induced cardiac hypertrophy, vascular inflammation and injury are attenuated by antioxidant supplementation in an animal model. *Front Pharmacol* 2016;7:397. doi:10.3389/fphar.2016.00397.
21. Zhang W, Lavine KJ, Epelman S, Evans SA, Weinheimer CJ, Barger PM, *et al.* Necrotic Myocardial Cells Release Damage-Associated Molecular Patterns That Provoke Fibroblast Activation *In Vitro* and Trigger Myocardial Inflammation and Fibrosis *In Vivo*. *J Am Heart Assoc* 2015;4:e001993. [doi: 10.1161/JAHA.115.001993].
22. Khudhur ZO, Smail SW, Awla HK, Ahmed GB, Khedhir YO, Amin K, *et al.* The effects of heavy smoking on oxidative stress, inflammatory biomarkers, vascular dysfunction, and hematological indices. *Sci Rep* 2025;15:18251. doi:10.1038/s41598-025-03075-8.
23. Khudhur ZO, Smail SW, Awla HK, Ahmed GB, Khedhir YO, Amin K, *et al.* The effects of heavy smoking on oxidative stress, inflammatory biomarkers, vascular dysfunction, and hematological indices. *Sci Rep* 2025;15:18251.
24. Li X, Xu H, Liu K, Shi M, Zeng X, Liu X. LXA4 alleviates inflammation and ferroptosis in cigarette smoke induced chronic obstructive pulmonary disease via the ALX/FPR2 receptor. *Int Immunopharmacol* 2025;151:114322.
25. Loffredo L, Maggio E, Bartimoccia S, Magna A, Totè CM, Bagnato C, *et al.* Tobacco smoke exposure and oxidative stress: The role of circulating lipopolysaccharides in heated and conventional products. *Antioxidants (Basel)* 2025;14:1316.
26. Cowling RT, Kupsy D, Kahn AM, Daniels LB, Greenberg BH. Mechanisms of Cardiac Collagen Deposition in Experimental Models and Human Disease. *Transl Res* 2019;209:138-55. doi: 10.1016/j.trsl.2019.03.004.
27. Wu JP, Chang-Lee SN, Day CH, Ho TJ, Viswanadha VP, Chung LC, *et al.* Secondhand smoke exposure enhances cardiac fibrosis effects on the aging rat hearts. *Acta Cardiol Sin* 2016;32:594-603. doi:10.6515/ACS20150824C.
28. Gajjar DN, Patel S, Shah DR, Shah GR, Patel AD, Patel VV. Comparative analysis of lipid profiles in young smokers and non-smokers. *Eur J Cardiovasc Med* 2025;15:396-9.
29. Copeland KM, Chen H, Chintapula U, Almasian M, Chung DK, Taylor AM, *et al.* Deteriorated biomechanical properties of human hypertrophied septum in response to cardiomyocyte enlargement, overexpressed collagen, and disarrayed microstructures. *Front Bioeng Biotechnol* 2025;13:1620594. doi:10.3389/fbioe.2025.1620594.
30. Rahman M, Alatiqi M, Al Jarallah M, Hussain MY, Monayem A, Panduranga P, *et al.* Cardiovascular effects of smoking and smoking cessation: A 2024 update. *Glob Heart* 2025;20:15. doi:10.5334/gh.1399.
31. Kwong JQ, Correll RN, Liu Q. Editorial: Cell death programs in the pathogenesis of heart disease. *Front Physiol* 2025;16:1581282. [doi: 10.3389/fphys.2025.1581282].
32. Chen Q, Zheng A, Xu X, Shi Z, Yang M, Sun S, *et al.* Nrf3-mediated mitochondrial superoxide promotes cardiomyocyte apoptosis and impairs cardiac functions by suppressing Pitx2. *Circulation* 2025;151:1024-46. doi:10.1161/CIRCULATIONAHA.124.070286.
33. Cheng HG, Noggle B, Flora JW. Comparative disease risks associated with cigarette smoking and use of moist smokeless tobacco and snus: An umbrella review of epidemiological evidence from the United States and Western Europe. *BMC Public Health* 2025;25:3765. [doi: 10.1186/s12889-025-23280-4].
34. Alsarhan AA, Al-Shawabkeh JD, Abu Laban NM, Ababneh SK, Shraideh Z, Badran D, *et al.* Ameliorating effects of Ascorbic acid and Ammi Visnaga seeds on cigarette and water- pipe smoking cytotoxicity in the lung and heart ventricle. *Braz J Biol* 2025;85:e290032.

Supplementary Table 1: Area of inflammation and lesion in the myocardium of rats in Group II (per animal)

Rat	Inflammation (μm^2)	Lesion (μm^2)
R1	3455.09	2786.44
R2	3812.75	1851.99
R3	6876.69	3305.50
R4	5032.87	1419.31
R5	7030.43	1032.87
Mean $\pm$ SD	5241.57 $\pm$ 1669.60	2079.22 $\pm$ 946.82
CV	31.85	45.54

Values for each rat represent the mean area of five randomly selected microscopic fields at 400 $\times$ magnification. Animal-level means ($n=5$) were used as the unit of statistical analysis. Group II (kretek smoke-exposed) rats received smoke exposure (1 kretek/rat/day) for 30 days. Raw data are available from the corresponding author upon reasonable request. SD: Standard deviation, CV: Coefficient of variation ($\text{SD}/\text{mean} \times 100\%$)

Supplementary Table 2: Myocardial collagen area fraction in Group II rats (aggregated per animal)

Rat	Collagen area fraction (%)
R1	21.40
R2	21.94
R3	22.28
R4	21.49
R5	22.80
Mean $\pm$ SD	21.98 $\pm$ 0.58

Values for each rat represent the mean collagen area fraction calculated from five randomly selected microscopic fields at 400 $\times$ magnification. Animal-level means ($n=5$) were used as the unit of statistical analysis. Group II (kretek smoke-exposed) rats received smoke exposure (1 kretek/rat/day) for 30 days. SD: Standard deviation

Supplementary Table 3: Comparison of region of interest area positive for p53 protein between Group I and Group II (aggregated per animal)

Rat	ROI area positive for p53 protein (μm^2)	
	Group I	Group II
1	215.93	233.29
2	205.06	219.19
3	204.29	226.26
4	193.33	222.72
5	180.75	221.93
Mean $\pm$ SD	199.87 $\pm$ 13.35	224.68 $\pm$ 5.44

ROI area positive for p53 protein represents the area of immunopositive p53 staining within the defined ROI. Values for each rat represent the mean calculated from five randomly selected microscopic fields per rat at 200 $\times$ magnification. Animal-level means ($n=5$ per group) were used as the unit of statistical analysis. Group I (control group) rats breathed normal air. Group II (kretek smoke-exposed group) rats received smoke exposure (1 kretek/rat/day) for 30 days. SD: Standard deviation, ROI: Region of interest